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- (71) Applicant (for all designated States except US): INCYTE GENOMICS, INC. [US/US]; 3160 Porter Drive, Palo Alto, CA 94304 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (for US only): CUNNINGHAM, Mary, Jane [US/US]; 1204 Manet Drive, Sunnyvale, CA 94087 (US). ZWEIGER, Gary, B. [US/US]; 765-10 San Antonio Road, Palo Alto, CA 94306 (US). KASER, Matthew, R. [GB/US]; 4793 Ewing Road, Castro Valley, CA 94546 (US). PANZER, Scott, R. [US/US]; 571 Bobolink Circle, Sunnyvale, CA 94087 (US). SEILHAMER, Jeffrey, J. [US/US]; 12555 La Cresta, Los Altos Hills, CA 94022 (US). YUE, Henry [US/US]; 826 Lois Avenue, Sunnyvale, CA 94087 (US). BAUGHN, Mariah, R. [US/US]; 14244 Santiago Road, San Leandro, CA 94577 (US). AZIMZAI, Yalda [US/US]; 5518 Boulder Canyon Drive, Castro Valley, CA 94552 (US). LAL, Preeti [IN/US]; 2382 Lass Drive, Santa Clara, CA 95054 (US).
- (74) Agents: TURNER, Christopher et al.; 3160 Porter Drive, Palo Alto, CA 94304 (US).
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(54) Title: MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

(57) Abstract: The present invention relates to mammalian nucleic acid and protein molecules comprising a plurality of nucleic acid and protein molecules. The mammalian nucleic acid molecules can be used as hybridizable array elements in a microarray in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action. The protein molecules can be used in a pharmaceutical composition. The present invention also relates to methods for screening compounds and therapeutics for metabolic responses indicative of a toxic compound or molecule.

MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

This application is filed under the Patent Cooperation Treaty and claims the benefit of U.S. Nonprovisional Application No. 09/443,184, our Docket No. PC-0007 US, filed 19th November, 1999.

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TECHNICAL FIELD

The present invention relates to mammalian nucleic acid and protein molecules, and methods for their use in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

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BACKGROUND ART

Toxicity testing is a mandatory and time-consuming part of drug development programs in the pharmaceutical industry. A more rapid screen to determine the effects upon metabolism and to detect toxicity of lead drug candidates may be the use of gene expression microarrays. For example, microarrays of various kinds may be produced using full length genes or gene fragments. These arrays can then be used to test samples treated with the drug candidates to elucidate the gene expression pattern associated with drug treatment. This gene pattern can be compared with gene expression patterns associated with compounds which produce known metabolic and toxicological responses.

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Benzo(a)pyrene is a known rodent and likely human carcinogen and is the prototype of a class of compounds, the polycyclic aromatic hydrocarbons (PAH). It is metabolized by several forms of cytochrome P450 (P450 isozymes) and associated enzymes to form both activated and detoxified metabolites. The ultimate metabolites are the bay-region diol epoxide, benzo(a)pyrene-7,8-diol-9,10-epoxide (BPDE) and the K-region diol epoxide, 9-hydroxy benzo(a)pyrene-4,5-oxide, both of which induce formation of DNA adducts. DNA adducts have been shown to persist in rat liver up to 56 days following treatment with benzo(a)pyrene at a dose of 10 mg/kg body weight three times per week for two weeks (Qu and Stacey (1996) Carcinogenesis 17:53-59).

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Acetaminophen is a widely-used analgesic. It is metabolized by specific cytochrome P450 isozymes with the majority of the drug undergoing detoxification by glucuronic acid, sulfate and glutathione conjugation pathways. However, at supratherapeutic doses, acetaminophen is metabolized to an active intermediate, *N*-acetyl-*p*-benzoquinone imine (NAPQI) which can cause hepatic and renal failure. NAPQI then binds to sulphydryl groups of proteins causing their inactivation and leading to subsequent cell death (Kroger *et al.* (1997) Gen. Pharmacol. 28:257-263).

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Clofibrate is an hypolidemic drug which lowers elevated levels of serum triglycerides. In rodents, chronic treatment produces hepatomegaly and an increase in hepatic peroxisomes (peroxisome

proliferation). Peroxisome proliferators (PPs) are a class of drugs which activate the PP-activated receptor in rodent liver, leading to enzyme induction, stimulation of S-phase, and a suppression of apoptosis (Hasmall and Roberts (1999) *Pharmacol. Ther.* 82:63-70). PPs include the fibrate class of hypolidemic drugs, phenobarbitone, thiazolidinediones, certain non-steroidal anti-inflammatory drugs, and naturally-occurring fatty acid-derived molecules (Gelman *et al.* (1999) *Cell. Mol. Life Sci.* 55:932-943). Clofibrate has been shown to increase levels of cytochrome P450 4A. It is also involved in transcription of β -oxidation genes as well as induction of PP-activated receptors (Kawashima *et al.* (1997) *Arch. Biochem. Biophys.* 347:148-154). Peroxisome proliferation that is induced by both clofibrate and the chemically-related compound fenofibrate is mediated by a common inhibitory effect on mitochondrial membrane depolarization (Zhou and Wallace (1999) *Toxicol. Sci.* 48:82-89).

Toxicological effects in the liver are also induced by other compounds. These can include carbon tetrachloride (a necrotic agent), hydrazine (a steatotic agent), α -naphthylisothiocyanate (a cholestatic agent), 4-acetylaminofluorene (a liver mitogen), and their corresponding metabolites, which are used in experimental protocols to measure toxicological responses (Waterfield *et al.* (1993) *Arch. Toxicol.* 67:244-254).

The present invention provides mammalian nucleic acid and protein molecules, their use in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

DISCLOSURE OF INVENTION

The invention provides a method for detecting or diagnosing the effect of a test compound or molecule associated with increased or decreased levels of nucleic acid molecules in a mammalian subject. The method comprises treating a mammalian subject with a known toxic compound or molecule which elicits a toxicological response, measuring levels of a plurality of nucleic acid molecules, selecting from the plurality of nucleic acid molecules those nucleic acid molecules that have levels modulated in samples treated with known toxic compounds or molecules when compared with untreated samples. Some of the levels may be upregulated by a toxic compound or molecule, others may be downregulated by a toxic compound or molecule, and still others may be upregulated with one known toxic compound or molecule and be downregulated with another known toxic compound or molecule. The selected nucleic acid molecules which are upregulated and downregulated by a known toxic compound or molecule are arrayed upon a substrate. The method further comprises measuring levels of nucleic acid molecules in the sample after the sample is treated with the toxic compound or molecule. Levels of nucleic acid molecules in a sample so treated are then compared with the plurality of the arrayed nucleic acid molecules to identify which sample nucleic acid molecules are upregulated and downregulated by the test

compound or molecule. In one embodiment, the nucleic acid molecules are hybridizable array elements of a microarray.

Preferably, the comparing comprises contacting the arrayed nucleic acid molecules with the sample nucleic acid molecules under conditions effective to form hybridization complexes between the arrayed nucleic acid molecules and the sample nucleic acid molecules; and detecting the presence or absence of the hybridization complexes. In this context, similarity may mean that at least 1, preferably at least 5, more preferably at least 10, of the upregulated arrayed nucleic acid molecules form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a greater extent than would the probes derived from a sample not treated with the test compound or molecule or a known toxic compound or molecule. Similarity may also mean that at least 1, preferably at least 5, more preferably at least 10, of the downregulated arrayed nucleic acid molecules form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a lesser extent than would the sample nucleic acid molecules of a sample not treated with the test compound or a known toxic compound. In one aspect, the arrayed nucleic acid molecules comprise SEQ ID NOs:1-47 or fragments thereof.

Preferred toxic compounds are selected from the group consisting of hypolipidemic drugs, n-alkylcarboxylic acids, n-alkylcarboxylic acid precursors, azole antifungal compounds, leukotriene D4 antagonists, herbicides, pesticides, phthalate esters, phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol and their corresponding metabolites, acetaminophen and its corresponding metabolites, benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like, carbon tetrachloride, hydrazine, α -naphthylisothiocyanate, 4-acetylaminofluorene, and their corresponding metabolites. Preferred tissues are selected from the group consisting of liver, kidney, brain, spleen, pancreas and lung.

The arrayed nucleic acid molecules comprise fragments of messenger RNA transcripts of genes that are upregulated-or-downregulated at least 2-fold, preferably at least 2.5-fold, more preferably at least 3-fold, in tissues treated with known toxic compounds when compared with untreated tissues. Preferred arrayed nucleic acid molecules are selected from the group consisting of SEQ ID NOs:1-47 or fragments thereof, some of whose expression is upregulated following treatment with a toxic compound or molecule and others of whose expression is downregulated following treatment with a toxic compound or molecule. More preferable are SEQ ID NOs:2, 4, 6, 8, 9, and 11 which are upregulated following treatment with a toxic compound or molecule, and SEQ ID NOs:1, 4, and 7 which are downregulated following treatment with a toxic compound or molecule.

The invention also provides a method comprising measuring levels of nucleic acid molecules in a sample after the sample is treated with a test compound or molecule. Levels of nucleic acid molecules in

a sample so treated are then compared with the plurality of the arrayed nucleic acid molecules to identify which sample nucleic acid molecules are upregulated and downregulated by the test compound or molecule. In one embodiment, the nucleic acid molecules are hybridizable array elements of a microarray.

5 Alternatively, the invention provides methods for screening a sample for a metabolic response to a test compound or molecule.

 Alternatively, the invention provides methods for screening a test compound or molecule for a previously unknown metabolic response.

10 In another aspect, the invention provides methods for preventing a toxicological response by administering complementary nucleotide molecules against one or more selected upregulated nucleic acid molecules or a ribozyme that specifically cleaves such molecules. Alternatively, a toxicological response may be prevented by administering sense nucleotide molecules for one or more selected downregulated nucleic acid molecules.

15 In yet another aspect, the invention provides methods for preventing a toxicological response by administering an agonist which initiates transcription of a gene comprising a downregulated nucleic acid molecule of the invention. Alternatively, a toxicological response may be prevented by administering an antagonist which prevents transcription of a gene comprising an upregulated nucleic acid molecule of the invention.

20 In another aspect, the invention provides nucleic acid molecules whose transcript levels are modulated in a sample during a metabolic response to a toxic compound or molecule. The invention also provides nucleic acid molecules whose transcript levels are upregulated in a sample during a metabolic response to a toxic compound or molecule. The invention also provides nucleic acid molecules whose transcript levels are downregulated in a sample during a metabolic response to a toxic compound or molecule. Upregulation or downregulation is at least 2-fold, more preferably at least 2.5-fold, even more preferably at least 3-fold. The metabolic response to a toxic compound or molecule may be a toxicological response. The invention also provides mammalian nucleic acid molecules which are homologous to the upregulated and downregulated nucleic acid molecules. In one aspect, preferred arrayed nucleic acid molecules are selected from the group consisting of SEQ ID NOs:1-47, or fragments thereof.

25 The invention also provides a method for using a molecule selected from SEQ ID NOs:1-59 or a portion thereof to screen a library of molecules to identify at least one ligand which specifically binds the selected molecule, the method comprising combining the selected molecule with the library of molecules under conditions allowing specific binding, and detecting specific binding, thereby identifying a ligand which specifically binds the selected molecule.

Such libraries include DNA and RNA molecules, peptides, peptide nucleic acids, agonists, antagonists, antibodies, immunoglobulins, drug compounds, pharmaceutical agents, and other ligands. In one aspect, the ligand identified using the method modulates the activity of the selected molecule. In an analogous method, the selected molecule or a portion thereof is used to purify a ligand. The method 5 involves combining the selected molecule or a portion thereof with a sample under conditions to allow specific binding, detecting specific binding between the selected molecule and ligand, recovering the bound selected molecule, and separating the selected molecule from the ligand to obtain purified ligand. The invention further provides a method for using at least a portion of the proteins encoded by SEQ ID NOs:1-47 and the proteins of SEQ ID NOs:48-59 to produce antibodies.

10 The invention further provides a method for inserting a marker gene into the genomic DNA of an animal to disrupt the expression of the natural nucleic acid molecule. The invention also provides a method for using the nucleic acid molecule to produce an animal model system, the method comprising constructing a vector containing the nucleic acid molecule; introducing the vector into a totipotent embryonic stem cell; selecting an embryonic stem cell with the vector integrated into genomic DNA; 15 microinjecting the selected cell into a blastocyst, thereby forming a chimeric blastocyst; transferring the chimeric blastocyst into a pseudopregnant dam, wherein the dam gives birth to a chimeric animal containing at least one additional copy of nucleic acid molecule in its germ line; and breeding the chimeric animal to generate a homozygous animal model system.

20 The invention also provides a substantially purified mammalian protein or a portion thereof. The invention further provides isolated and purified proteins encoded by the nucleic acid molecules of SEQ ID NOs:1-11, 17-33, 36, 39, and 41. The invention further provides isolated and purified protein molecule of SEQ ID NOs:50 and 53. Additionally, the invention provides a pharmaceutical composition comprising a substantially purified mammalian protein or a portion thereof in conjunction with a pharmaceutical carrier.

25 The invention further provides an isolated and purified mammalian nucleic acid molecule variant having at least 70% nucleic acid sequence identity to the mammalian nucleic acid molecule selected from SEQ ID NO:1-47 and fragments thereof. The invention also provides an isolated and purified nucleic acid molecule having a sequence which is complementary to the mammalian nucleic acid molecule comprising a nucleic acid molecule selected from SEQ ID NO:1-47 and fragments thereof.

30 The invention further provides an expression vector containing at least a fragment of the mammalian nucleic acid molecule selected from the group consisting of SEQ ID NOs:1-47. In another aspect, the expression vector is contained within a host cell.

The invention also provides a method for producing a mammalian protein, the method comprising the steps of: (a) culturing the host cell containing an expression vector containing a

mammalian nucleic acid molecule of the invention under conditions suitable for the expression of the polypeptide; and (b) recovering the polypeptide from the host cell culture.

The invention also provides a pharmaceutical composition comprising a substantially purified mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and the amino acid sequence of SEQ ID NOs:50 and 53 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

5 The invention further includes an isolated and purified antibody which binds to a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53 or fragments thereof. The invention also provides a purified agonist and a purified antagonist.

10

BRIEF DESCRIPTION OF THE SEQUENCE LISTING

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The Sequence Listing contains the nucleic acid sequence of exemplary mammalian nucleic acid molecules of the invention, SEQ ID NOs:1-47, 60-135, 137, and 138; the protein sequence of exemplary mammalian protein molecules of the invention, SEQ ID NOs:48-59 and 136.

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MODES FOR CARRYING OUT THE INVENTION

Definitions

“Sample” is used in its broadest sense. A sample containing nucleic acid molecules may comprise a bodily fluid; a cell; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; genomic DNA, RNA, or cDNA in solution or bound to a substrate; a biological tissue or biopsy thereof; a fingerprint or tissue print; natural or synthetic fibres; in a solution; in a liquid suspension; in a gaseous suspension; in an aerosol; and the like.

“Plurality” refers preferably to a group of one or more members, preferably to a group of at least about 10, and more preferably to a group of at least about 100 members, and even more preferably a group of 10,000 members.

“Substrate” refers to a rigid or semi-rigid support to which nucleic acid molecules or proteins are bound and includes membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, capillaries or other tubing, plates, polymers, and microparticles with a variety of surface forms including wells, trenches, pins, channels and pores.

“Modulates” refers to a change in activity (biological, chemical, or immunological) or lifespan resulting from specific binding between a molecule and either a nucleic acid molecule or a protein.

“Microarray” refers to an ordered arrangement of hybridizable array elements on a substrate. The array elements are arranged so that there are preferably at least ten or more different array elements, more preferably at least 100 array elements, even more preferably at least 1000 array elements, and most preferably 10,000. Furthermore, the hybridization signal from each of the array elements is individually distinguishable. In a preferred embodiment, the array elements comprise nucleic acid molecules.

5 “Nucleic acid molecule” refers to a nucleic acid, oligonucleotide, nucleotide, polynucleotide or any fragment thereof. It may be DNA or RNA of genomic or synthetic origin, double-stranded or single-stranded, and combined with carbohydrate, lipids, protein, or other materials to perform a particular activity such as transformation or form a useful composition such as a peptide nucleic acid (PNA). “Oligonucleotide” is substantially equivalent to the terms amplimer, primer, oligomer, element, target, and probe and is preferably single stranded.

10 “Protein” refers to an amino acid sequence, oligopeptide, peptide, polypeptide, or portions thereof whether naturally occurring or synthetic. Exemplary portions are the first twenty consecutive amino acids of a mammalian protein encoded by SEQ ID NOS:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOS:50 and 53.

15 “Up-regulated” refers to a nucleic acid molecule whose levels increased in a treated sample compared with the nucleic acid molecule in an untreated sample.

“Down-regulated” refers to nucleic acid molecule whose levels decreased in a treated sample compared with the nucleic acid molecule in an untreated sample.

20 “Toxic compound” or “toxic agent” is any compound, molecule, or agent that elicits a biochemical, metabolic, and physiological response in an individual or animal, such as i) DNA damage, ii) cell damage, iii) organ damage or cell death, or iv) clinical morbidity or mortality.

“Toxicological response” refers to a biochemical, metabolic, and physiological response in an individual or animal which has been exposed to a toxic compound or agent.

25 “Fragment” refers to an Incyte clone or any part of a molecule which retains a usable, functional characteristic. Useful fragments include oligonucleotides and polynucleotides which may be used in hybridization or amplification technologies or in regulation of replication, transcription or translation. Exemplary fragments are the first sixty consecutive nucleotides of SEQ ID NOS:1-47. Useful fragments also include polypeptides and protein molecules which have antigenic potential and which may be used 30 with a suitable pharmaceutical carrier in a pharmaceutical composition. Exemplary fragments are the first twenty consecutive amino acids of a mammalian protein encoded by SEQ ID NOS:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOS:50 and 53.

“Hybridization complex” refers to a complex between two nucleic acid molecules by virtue of the formation of hydrogen bonds between purines and pyrimidines.

"Ligand" refers to any compound, molecule, or agent which will bind specifically to a complementary site on a nucleic acid molecule or protein. Such ligands stabilize or modulate the activity of nucleic acid molecules or proteins of the invention and may be composed of at least one of the following: inorganic and organic substances including nucleic acids, proteins, carbohydrates, fats, and lipids.

"Percent identity" or "% identity" refers to the percentage of sequence similarity found in a comparison of two or more amino acid or nucleic acid sequences. Percent identity can be determined electronically, e.g., by using the MEGALIGN program (DNASTAR, Madison WI) which creates alignments between two or more sequences according to methods selected by the user, e.g., the clustal method. (See, e.g., Higgins, D.G. and P.M. Sharp (1988) Gene 73:237-244.) The clustal algorithm groups sequences into clusters by examining the distances between all pairs. The clusters are aligned pairwise and then in groups. The percentage similarity between two amino acid sequences, e.g., sequence A and sequence B, is calculated by dividing the length of sequence A, minus the number of gap residues in sequence A, minus the number of gap residues in sequence B, into the sum of the residue matches between sequence A and sequence B, times one hundred. Gaps of low or of no similarity between the two amino acid sequences are not included in determining percentage similarity. Percent identity between nucleic acid sequences can also be counted or calculated by other methods known in the art, e.g., the Jotun Hein method. (See, e.g., Hein, J. (1990) Methods Enzymol. 183:626-645.) Identity between sequences can also be determined by other methods known in the art, e.g., by varying hybridization conditions.

"Substantially purified" refers to nucleic acid molecules or proteins that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably about 75% free, and most preferably about 90% free, from other components with which they are naturally associated.

25 The Invention

The present invention provides mammalian nucleic acid and protein molecules and method of using the nucleic acid molecules for screening test compounds and molecules for toxicological responses. Additionally the invention provides methods for characterizing the toxicological responses of a sample to a test compound or molecule. In particular, the present invention provides a composition comprising a plurality of nucleic acid molecules derived from human cDNA libraries, monkey cDNA libraries, mouse cDNA libraries, normal rat liver cDNA libraries, normalized rat liver cDNA libraries, prehybridized rat liver cDNA libraries, subtracted rat liver cDNA libraries, and rat kidney cDNA libraries. The nucleic acid molecules have been further selected for exhibiting upregulated or downregulated gene expression

in rat livers when the rats have been exposed to a known hepatotoxin, including a peroxisomal proliferator (PP), acetaminophen or one of its corresponding metabolites, a polycyclic aromatic hydrocarbon (PAH), carbon tetrachloride, hydrazine, α -naphthylisothiocyanate, 4-acetylaminofluorene, and their corresponding metabolites.

5 PPs include hypolipidemic drugs, such as clofibrate, fenofibrate, clofenic acid, nafenopin, gemfibrozil, ciprofibrate, bezafibrate, halofenate, simfibrate, benzofibrate, etofibrate, WY-14,643, and the like; n-alkylcarboxylic acids, such as trichloroacetic acid, valproic acid, hexanoic acid, and the like; n-alkylcarboxylic acid precursors, such as trichloroethylene, tetrachloroethylene, and the like; azole antifungal compounds, such as bifonazole, and the like; leukotriene D4 antagonists; herbicides; 10 pesticides; phthalate esters, such as di-[2-ethylhexyl] phthalate, mono-[2-ethylhexyl] phthalate, and the like; and natural chemicals, such as phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol, and the like. In a preferred embodiment the toxin is clofibrate, or one of its corresponding metabolites. In another preferred embodiment the toxin is fenofibrate, or one of its corresponding metabolites.

15 PAHs include compounds such as benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like. In a preferred embodiment the toxin is benzo(a)pyrene, or one of its corresponding metabolites.

SEQ ID NOs:1-16 were identified by their pattern of at least two-fold upregulation or downregulation following hybridization with sample nucleic acid molecules from rat liver tissue treated 20 with a known toxic compound. SEQ ID NOs:17-47 were identified by their homology to the sample nucleic acid molecules from rat liver tissue treated with a known toxic compound. These and other nucleic acid molecules can be immobilized on a substrate as hybridizable array elements in a microarray format. The microarray may be used to characterize gene expression patterns associated with novel compounds to elucidate any toxicological responses or to monitor the effects of treatments during clinical trials or therapy where metabolic responses to toxic compounds may be expected.

When the nucleic acid molecules are employed as hybridizable array elements in a microarray, the array elements are organized in an ordered fashion so that each element is present at a specified location on the substrate. Because the array elements are at specified locations on the substrate, the hybridization patterns and intensities (which together create a unique expression profile) can be 30 interpreted in terms of expression levels of particular genes and can be correlated with a toxicological response associated with a test compound or molecule.

The invention also provides a substantially purified and isolated mammalian protein comprising the protein molecule of SEQ ID NOs:50 and 53 or portion thereof. The invention further provides isolated and purified proteins encoded by the nucleic acid molecules of SEQ ID NOs:1-11, 17-33, 36, 39,

and 41, or portion thereof.

Furthermore, the present invention provides methods for screening test compounds or therapeutics for potential toxicological responses and for screening a sample's toxicological response to a particular test compound or molecule. Briefly, these methods entail treating a sample with the test compound or molecule to elicit a change in gene expression patterns comprising the expression of a plurality of sample nucleic acid molecules. Nucleic acid molecules are selected by identifying those genes in rat liver or kidney that are upregulated-or-downregulated at least 2-fold, more preferably at least 2.5-fold, most preferably at least 3-fold, when treated with a known toxic compound or molecule. The nucleic acid molecules are arrayed on a substrate. Then, the arrayed nucleic acid molecules and sample nucleic acid molecules are combined under conditions effective to form hybridization complexes which may be detected by methods well known in the art. Detection of higher or lower levels of such hybridization complexes compared with hybridization complexes derived from untreated samples and samples treated with a compound that is known not to induce a toxicological response correlates with a toxicological response of a test compound or a toxicological response to a molecule.

Complementary DNA libraries

Molecules are identified that reflect all or most of the genes that are expressed in rat liver or kidney. Molecules may be identified by isolating clones derived from several types of rat cDNA libraries, including normal rat cDNA libraries, normalized rat cDNA libraries, prehybridized rat cDNA libraries, and subtracted cDNA libraries. Clone inserts derived from these clones may be partially sequenced to generate expressed sequence tags (ESTs). Molecules are also identified by comparing the clones from rat cDNA libraries with clones from human, monkey, and mouse cDNA libraries using computer software nucleic acid comparison programs such as BLAST (see, e.g., Altschul, S.F. (1993) J. Mol. Evol. 3:290-300; Altschul, *et al.* (1990) J. Mol. Biol. 215:403-410).

In one embodiment, two collections of ESTs are identified and sequenced. A first collection of ESTs (the originator molecules) are derived from rat liver and kidney and are derived from the cDNA libraries presented in the Examples. A second collection includes ESTs derived from other rat cDNA libraries available in the ZOOSEQ database (Incyte Pharmaceuticals, Inc. Palo Alto CA).

The two collections of ESTs are clustered electronically to form master clusters of ESTs. Master clusters are formed by identifying overlapping EST molecules and assembling these ESTs. A nucleic acid fragment assembly tool, such as the Phrap tool (Phil Green, University of Washington) and the GELVIEW fragment assembly system (GCG, Madison WI), can be used for this purpose. The minimum number of clones which constitute a cluster is two. In another embodiment, a collection of human genes known to be expressed in response to toxic agents are used to select representative ESTs from the 113 rat cDNA libraries. The master cluster process is repeated for these molecules.

After assembling the clustered consensus nucleic acid sequences, a representative 5' clone is nominated from each master cluster. The most 5' clone is preferred because it is most likely to contain the complete gene. The nomination process is described in greater detail in "Relational Database and System for Storing Information Relating to Biomolecular Sequences and Reagents", USSN 09/034,807, filed March 4, 1998, herein incorporated in its entirety by reference. The EST molecules are used as array elements on a microarray.

Selection of arrayed nucleic acid molecules

Samples are treated, preferably at subchronic doses, with one or more known toxic compounds over a defined time course. Preferably, the agents are peroxisomal proliferators (PPs), acetaminophen or one of its corresponding metabolites, polycyclic aromatic hydrocarbons (PAHs), carbon tetrachloride, hydrazine, α -naphthylisothiocyanate, 4-acetylaminofluorene, or their corresponding metabolites.

The gene expression patterns derived from such treated biological samples can be compared with the gene expression patterns derived from untreated biological samples to identify and select nucleic acid molecules whose expression is either upregulated or downregulated due to the response to the toxic compounds. These selected molecules may then be employed as array elements alone or in combination with other array element molecules. Such a microarray is particularly useful to detect and characterize gene expression patterns associated with known toxic compounds. Such gene expression patterns can then be used for comparison to identify other compounds which also elicit a toxicological response.

The arrayed nucleic acid molecules can be manipulated to optimize their performance in hybridization. To optimize hybridization, the arrayed nucleic acid molecules are examined using a computer algorithm to identify portions of genes without potential secondary structure. Such computer algorithms are well known in the art and are part of OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or LASERGENE software (DNASTAR, Madison WI). These programs can search within nucleic acid sequences to identify stem loop structures and tandem repeats and to analyze G + C content of the sequence (those molecules with a G + C content greater than 60% are excluded). Alternatively, the arrayed nucleic acid molecules can be optimized by trial and error. Experiments can be performed to determine whether sample nucleic acid molecules and complementary arrayed nucleic acid molecules hybridize optimally under experimental conditions.

The arrayed nucleic acid molecules can be any RNA-like or DNA-like material, such as mRNAs, cDNAs, genomic DNA, peptide nucleic acids, branched DNAs and the like. The arrayed nucleic acid molecules can be in sense or antisense orientations.

In one embodiment, the arrayed nucleic acid molecules are cDNAs. The size of the DNA sequence of interest may vary, and is preferably from 50 to 10,000 nucleotides, more preferably from 150 to 3,500 nucleotides. In a second embodiment, the nucleic acid molecules are vector DNAs. In this case

the size of the DNA sequence of interest, i.e., the insert sequence, may vary from about 50 to 10,000 nucleotides, more preferably from about 150 to 3,500 nucleotides.

The nucleic acid molecule sequences of the Sequence Listing have been prepared by current, state-of-the-art, automated methods and, as such, may contain occasional sequencing errors and unidentified nucleotides. Nucleotide analogues can be incorporated into the nucleic acid molecules by methods well known in the art. The only requirement is that the incorporated nucleotide analogues must serve to base pair with sample nucleic acid molecules. For example, certain guanine nucleotides can be substituted with hypoxanthine which base pairs with cytosine residues. However, these base pairs are less stable than those between guanine and cytosine. Alternatively, adenine nucleotides can be substituted with 2,6-diaminopurine which can form stronger base pairs than those between adenine and thymidine. Additionally, the nucleic acid molecules can include nucleotides that have been derivatized chemically or enzymatically. Typical modifications include derivatization with acyl, alkyl, aryl or amino groups.

The nucleic acid molecules can be immobilized on a substrate via chemical bonding.

Furthermore, the molecules do not have to be directly bound to the substrate, but rather can be bound to the substrate through a linker group. The linker groups are typically about 6 to 50 atoms long to provide exposure to the bound nucleic acid molecule. Preferred linker groups include ethylene glycol oligomers, diamines, diacids and the like. Reactive groups on the substrate surface react with one of the terminal portions of the linker to bind the linker to the substrate. The other terminal portion of the linker is then functionalized for binding the nucleic acid molecule. Preferred substrates are any suitable rigid or semirigid support, including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which the arrayed nucleic acid molecules are bound.

The samples can be any sample comprising sample nucleic acid molecules and obtained from any bodily fluid (blood, urine, saliva, phlegm, gastric juices, etc.), cultured cells, biopsies, or other tissue preparations. The samples can be derived from any species, but preferably from eukaryotic species, and more preferably from mammalian species such as rat and human.

DNA or RNA can be isolated from the sample according to any of a number of methods well known to those of skill in the art. For example, methods of purification of nucleic acids are described in Tijssen, P. (1993) Laboratory Techniques in Biochemistry and Molecular Biology: Hybridization With Nucleic Acid Probes, Part I. Theory and Nucleic Acid Preparation, Elsevier, New York, NY. In one preferred embodiment, total RNA is isolated using the TRIZOL total RNA isolation reagent (Life Technologies, Inc., Gaithersburg MD) and mRNA is isolated using oligo d(T) column chromatography or

glass beads. When sample nucleic acid molecules are amplified it is desirable to amplify the sample nucleic acid molecules and maintain the relative abundances of the original sample, including low abundance transcripts. RNA can be amplified in vitro, in situ, or in vivo (See Eberwine US Patent No. 5,514,545).

5 It is also advantageous to include controls within the sample to assure that amplification and labeling procedures do not change the true distribution of nucleic acid molecules in a sample. For this purpose, a sample is spiked with an amount of a control nucleic acid molecule predetermined to be detectable upon hybridization to its complementary arrayed nucleic acid molecule and the composition of nucleic acid molecules includes reference nucleic acid molecules which specifically hybridize with the
10 control arrayed nucleic acid molecules. After hybridization and processing, the hybridization signals obtained should reflect accurately the amounts of control arrayed nucleic acid molecules added to the sample.

Prior to hybridization, it may be desirable to fragment the sample nucleic acid molecules. Fragmentation improves hybridization by minimizing secondary structure and cross-hybridization to
15 other sample nucleic acid molecules in the sample or noncomplementary nucleic acid molecules. Fragmentation can be performed by mechanical or chemical means.

Labeling

The sample nucleic acid molecules may be labeled with one or more labeling moieties to allow for detection of hybridized arrayed/sample nucleic acid molecule complexes. The labeling moieties can
20 include compositions that can be detected by spectroscopic, photochemical, biochemical, bioelectronic, immunochemical, electrical, optical or chemical means. The labeling moieties include radioisotopes, such as ^{32}P , ^{33}P or ^{35}S , chemiluminescent compounds, labeled binding proteins, heavy metal atoms, spectroscopic markers, such as fluorescent markers and dyes, magnetic labels, linked enzymes, mass spectrometry tags, spin labels, electron transfer donors and acceptors, and the like. Preferred fluorescent
25 markers include Cy3 and Cy5 fluorophores (Amersham Pharmacia Biotech, Piscataway NJ).

Hybridization

The nucleic acid molecule sequence of SEQ ID NOS:1-47 and fragments thereof can be used in various hybridization technologies for various purposes. Hybridization probes may be designed or derived from SEQ ID NOS:1-47. Such probes may be made from a highly specific region such as the 5' regulatory region or from a conserved motif, and used in protocols to identify naturally occurring sequences encoding the mammalian protein, allelic variants, or related sequences, and should preferably have at least 50% sequence identity to any of the protein sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NOS:1-47 or from genomic sequences including promoters, enhancers, and introns of the mammalian gene.

Hybridization or PCR probes may be produced using oligolabeling, nick translation, end-labeling, or PCR amplification in the presence of the labeled nucleotide. A vector containing the nucleic acid sequence may be used to produce an mRNA probe in vitro by addition of an RNA polymerase and labeled nucleic acid molecules. These procedures may be conducted using commercially available kits such as those provided by Amersham Pharmacia Biotech.

The stringency of hybridization is determined by G+C content of the probe, salt concentration, and temperature. In particular, stringency can be increased by reducing the concentration of salt or raising the hybridization temperature. In solutions used for some membrane based hybridizations, additions of an organic solvent such as formamide allows the reaction to occur at a lower temperature.

Hybridization can be performed at low stringency with buffers, such as 5 x SSC with 1% sodium dodecyl sulfate (SDS) at 60°C, which permits the formation of a hybridization complex between nucleotide sequences that contain some mismatches. Subsequent washes are performed at higher stringency with buffers such as 0.2 x SSC with 0.1% SDS at either 45°C (medium stringency) or 68°C (high stringency). At high stringency, hybridization complexes will remain stable only where the nucleic acid sequences are completely complementary. In some membrane-based hybridizations, preferably 35% or most preferably 50%, formamide can be added to the hybridization solution to reduce the temperature at which hybridization is performed, and background signals can be reduced by the use of other detergents such as Sarkosyl or Triton X-100 and a blocking agent such as salmon sperm DNA. Selection of components and conditions for hybridization are well known to those skilled in the art and are reviewed in Ausubel (supra) and Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY.

Hybridization specificity can be evaluated by comparing the hybridization of specificity-control nucleic acid molecules to specificity-control sample nucleic acid molecules that are added to a sample in a known amount. The specificity-control arrayed nucleic acid molecules may have one or more sequence mismatches compared with the corresponding arrayed nucleic acid molecules. In this manner, whether only complementary arrayed nucleic acid molecules are hybridizing to the sample nucleic acid molecules or whether mismatched hybrid duplexes are forming is determined.

Hybridization reactions can be performed in absolute or differential hybridization formats. In the absolute hybridization format, nucleic acid molecules from one sample are hybridized to the molecules in a microarray format and signals detected after hybridization complex formation correlate to nucleic acid molecule levels in a sample. In the differential hybridization format, the differential expression of a set of genes in two biological samples is analyzed. For differential hybridization, nucleic acid molecules from both biological samples are prepared and labeled with different labeling moieties. A mixture of the two labeled nucleic acid molecules is added to a microarray. The microarray is then examined under

conditions in which the emissions from the two different labels are individually detectable. Molecules in the microarray that are hybridized to substantially equal numbers of nucleic acid molecules derived from both biological samples give a distinct combined fluorescence (Shalon et al. PCT publication WO95/35505). In a preferred embodiment, the labels are fluorescent markers with distinguishable emission spectra, such as Cy3 and Cy5 fluorophores.

After hybridization, the microarray is washed to remove nonhybridized nucleic acid molecules and complex formation between the hybridizable array elements and the nucleic acid molecules is detected. Methods for detecting complex formation are well known to those skilled in the art. In a preferred embodiment, the nucleic acid molecules are labeled with a fluorescent label and measurement of levels and patterns of fluorescence indicative of complex formation is accomplished by fluorescence microscopy, preferably confocal fluorescence microscopy.

In a differential hybridization experiment, nucleic acid molecules from two or more different biological samples are labeled with two or more different fluorescent labels with different emission wavelengths. Fluorescent signals are detected separately with different photomultipliers set to detect specific wavelengths. The relative abundances/expression levels of the nucleic acid molecules in two or more samples is obtained.

Typically, microarray fluorescence intensities can be normalized to take into account variations in hybridization intensities when more than one microarray is used under similar test conditions. In a preferred embodiment, individual arrayed-sample nucleic acid molecule complex hybridization intensities are normalized using the intensities derived from internal normalization controls contained on each microarray.

The labeled sample emits specific wavelengths which are detected using a plurality of photomultipliers. The nucleic acid molecules whose relative abundance/expression levels are modulated by treatment of a sample with a known toxic compound can be used as hybridizable elements in a microarray. Such a microarray can be employed to identify expression profiles associated with particular toxicological responses. Then, a particular subset of these photomultipliers set to detect specific wavelengths. The relative expression levels of the arrayed nucleic acid molecules can be identified as to which arrayed nucleic acid molecule expression is modulated in response to a particular toxicological agent. These photomultipliers are set to detect specific wavelengths. The relative expression levels of the nucleic acid molecules can be employed to identify other compounds with a similar toxicological response.

Alternatively, for some treatments with known side effects, the microarray, and expression patterns derived therefrom, is employed to prospectively define the treatment regimen. A dosage is established that minimizes expression patterns associated with undesirable side effects. This approach

may be more sensitive and rapid than waiting for the patient to show toxicological side effects before altering the course of treatment.

Generally, the method for screening a library of test compounds or molecules to identify those with a toxicological response entails selecting a plurality of arrayed genes whose expression levels are modulated in tissues treated with known toxic compounds when compared with untreated tissues. Then a sample is treated with the test compound or molecule to induce a pattern of gene expression comprising the expression of a plurality of sample nucleic acid molecules. Tissues from a mammalian subject treated at various dosages of the test compound may be screened to determine which doses may be toxic.

Then, the expression levels of the arrayed genes and the sample nucleic acid molecules are compared to identify those compounds that induce expression levels of the sample nucleic acid molecules that are similar to those of the arrayed genes. In one preferred embodiment, gene expression levels are compared by contacting the arrayed genes with the sample nucleic acid molecules under conditions effective to form hybridization complexes between arrayed genes and sample nucleic acid molecules; and detecting the presence or absence of the hybridization complexes.

Similarity may mean that at least 1, preferably at least 5, more preferably at least 10, of the upregulated arrayed genes form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a greater extent than would the nucleic acid molecules of a sample not treated with the test compound. Similarity may also mean that at least 1, preferably at least 5, more preferably at least 10, of the downregulated nucleic acid molecules form hybridization complexes with the arrayed genes at least once during a time course to a lesser extent than would the nucleic acid molecules of a sample not treated with the test compound.

Such a similarity of expression patterns means that a toxicological response is associated with the compound or therapeutic tested. Preferably, the toxic compounds belong to the class of peroxisomal proliferators (PPs), including hypolipidemic drugs, such as clofibrate, fenofibrate, clofenic acid, nafenopin, gemfibrozil, ciprofibrate, bezafibrate, halofenate, simfibrate, benzofibrate, etofibrate, WY-14,643, and the like; n-alkylcarboxylic acids, such as trichloroacetic acid, valproic acid, hexanoic acid, and the like; n-alkylcarboxylic acid precursors, such as trichloroethylene, tetrachloroethylene, and the like; azole antifungal compounds, such as bifonazole, and the like; leukotriene D4 antagonists; herbicides; pesticides; phthalate esters, such as di-[2-ethylhexyl] phthalate, mono-[2-ethylhexyl] phthalate, and the like; and natural chemicals, such as phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol, and the like. In another embodiment, the toxic compound is acetaminophen or one of its corresponding metabolites. In yet another embodiment, the toxic compounds are polycyclic aromatic hydrocarbons (PAHs), including compounds such as benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like. Of

particular interest is the study of the toxicological responses of these compounds on the liver, kidney, brain, spleen, pancreas, and lung.

Modification of Gene Expression Using Nucleic Acids

Gene expression may be modified by designing complementary or antisense molecules (DNA, RNA, or PNA) to the control, 5', 3', or other regulatory regions of the mammalian gene.

Oligonucleotides designed with reference to the transcription initiation site are preferred. Similarly, inhibition can be achieved using triple helix base-pairing which inhibits the binding of polymerases, transcription factors, or regulatory molecules (Gee *et al.* In: Huber and Carr (1994) Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary molecule may also be designed to block translation by preventing binding between ribosomes and mRNA. In one alternative, a library of nucleic acid molecules or fragments thereof may be screened to identify those which specifically bind a regulatory, nontranslated sequence .

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA followed by endonucleolytic cleavage at sites such as GUA, GUU, and GUC. Once such sites are identified, an oligonucleotide with the same sequence may be evaluated for secondary structural features which would render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing their hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary nucleic acids and ribozymes of the invention may be prepared via recombinant expression, *in vitro* or *in vivo*, or using solid phase phosphoramidite chemical synthesis. In addition, RNA molecules may be modified to increase intracellular stability and half-life by addition of flanking sequences at the 5' and/or 3' ends of the molecule or by the use of phosphorothioate or 2' O-methyl rather than phosphodiester linkages within the backbone of the molecule. Modification is inherent in the production of PNAs and can be extended to other nucleic acid molecules. Either the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, and or the modification of adenine, cytidine, guanine, thymine, and uridine with acetyl-, methyl-, thio- groups renders the molecule less available to endogenous endonucleases.

Screening Assays

The nucleic acid molecule encoding the mammalian protein may be used to screen a library of molecules for specific binding affinity. The libraries may be DNA molecules, RNA molecules, PNAs, peptides, proteins such as transcription factors, enhancers, repressors, and other ligands which regulate the activity, replication, transcription, or translation of the nucleic acid molecule in the biological system. The assay involves combining the mammalian nucleic acid molecule or a fragment thereof with the

library of molecules under conditions allowing specific binding, and detecting specific binding to identify at least one molecule which specifically binds the nucleic acid molecule.

Similarly the mammalian protein or a portion thereof may be used to screen libraries of molecules in any of a variety of screening assays. The portion of the protein employed in such screening may be free in solution, affixed to an abiotic or biotic substrate (e.g. borne on a cell surface), or located intracellularly. Specific binding between the protein and molecule may be measured. Depending on the kind of library being screened, the assay may be used to identify DNA, RNA, or PNA molecules, agonists, antagonists, antibodies, immunoglobulins, inhibitors, peptides, proteins, drugs, or any other ligand, which specifically binds the protein. One method for high throughput screening using very small assay volumes and very small amounts of test compound is described in USPN 5,876,946, incorporated herein by reference, which screens large numbers of molecules for enzyme inhibition or receptor binding.

Purification of Ligand

The nucleic acid molecule or a fragment thereof may be used to purify a ligand from a sample. A method for using a mammalian nucleic acid molecule or a fragment thereof to purify a ligand would involve combining the nucleic acid molecule or a fragment thereof with a sample under conditions to allow specific binding, detecting specific binding, recovering the bound protein, and using an appropriate agent to separate the nucleic acid molecule from the purified ligand.

Similarly, the protein or a portion thereof may be used to purify a ligand from a sample. A method for using a mammalian protein or a portion thereof to purify a ligand would involve combining the protein or a portion thereof with a sample under conditions to allow specific binding, detecting specific binding between the protein and ligand, recovering the bound ligand, and using an appropriate chaotropic agent to separate the protein from the purified ligand.

Pharmacology

Pharmaceutical compositions are those substances wherein the active ingredients are contained in an effective amount to achieve a desired and intended purpose. The determination of an effective dose is well within the capability of those skilled in the art. For any compound, the therapeutically effective dose may be estimated initially either in cell culture assays or in animal models. The animal model is also used to achieve a desirable concentration range and route of administration. Such information may then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of protein or inhibitor which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity of such agents may be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., ED₅₀ (the dose therapeutically effective in 50% of the population) and LD₅₀ (the dose lethal to 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index, and it may be expressed as

the ratio, LD₅₀/ED₅₀. Pharmaceutical compositions which exhibit large therapeutic indexes are preferred. The data obtained from cell culture assays and animal studies are used in formulating a range of dosage for human use.

5 MODEL SYSTEMS

Animal models may be used as bioassays where they exhibit a toxic response similar to that of humans and where exposure conditions are relevant to human exposures. Mammals are the most common models, and most toxicity studies are performed on rodents such as rats or mice because of low cost, availability, and abundant reference toxicology. Inbred or outbred rodent strains provide a convenient model for investigation of the physiological consequences of under- or over-expression of genes of interest and for the development of methods for diagnosis and treatment of diseases. A mammal inbred to over-express a particular gene, so that the protein is secreted in milk, may also serve as a convenient source of the protein expressed by that gene.

Toxicology

15 Toxicology is the study of the effects of test compounds, molecules, or toxic agents on living systems to identify adverse effects. The majority of toxicity studies are performed on rats or mice to help predict whether adverse effects of agents will occur in humans. Observation of qualitative and quantitative changes in physiology, behavior, homeostatic, developmental, and reproductive processes, and lethality are used to generate profiles of safe or toxic responses and to assess the consequences on 20 human health following exposure to the agent.

Toxicological tests measure the effects of a single, repeated, or long-term exposure of a subject to a substance. Substances may be tested for specific endpoints such as cytotoxicity, mutagenicity, carcinogenicity and teratogenicity. Degree of response varies according to the route of exposure (contact, ingestion, injection, or inhalation), age, sex, genetic makeup, and health status of the subject. Other tests 25 establish the toxicokinetic and toxicodynamic properties of substances. Toxicokinetic studies trace the absorption, distribution in subject tissues, metabolism, storage, and excretion of substances. Toxicodynamic studies chart biological responses that are consequences of the presence of the substance in the subject tissues.

Genetic toxicology identifies and analyzes the ability of an agent to produce damage at a cellular 30 or subcellular level. Such genotoxic agents usually have common chemical or physical properties that facilitate interaction with nucleic acids and are most harmful when mutated chromosomes are passed along to progeny. Toxicological studies may identify agents that increase the frequency of structural or functional abnormalities in progeny if administered to either parent before conception, to the mother during pregnancy, or to the developing organism. Mice and rats are most frequently used in these tests

because of their short reproductive cycle which allows investigators to breed sufficient quantities of individual animals to satisfy statistical requirements.

All types of toxicology studies on experimental animals involve preparation of a suitable form of the compound for administration, selection of the route of administration, and selection of a species which resembles the species of pharmacological interest. Dose concentrations of the compound are varied to identify, measure, and investigate a range of dose-related effects related to exposure.

Acute toxicity tests are based on a single administration of the agent to the subject to determine the symptomology or lethality of the agent. Three experiments are conducted; an experiment to define the initial dose range; an experiment to narrow the range of effective doses; and a final experiment to establish the dose-response curve.

Prolonged and subchronic toxicity tests are based on the repeated administration of the agent. Rat and dog are commonly used in these studies to provide data from species in different taxonomic orders. With the exception of carcinogenesis, there is considerable evidence that daily administration of an agent at high-dose concentrations for periods of three to four months will reveal most forms of toxicity in adult animals.

Chronic toxicity tests, with a duration of a year or more, are used to demonstrate either the absence of toxicity or the carcinogenic potential of an agent. When studies are conducted on rats, a minimum of at least one test group plus one control group are used. Animals are quarantined, examined for health, and monitored at the outset and at intervals throughout the experiment.

20 Transgenic Animal Models

Transgenic rodents which over-express or under-express a gene of interest may be inbred and used to model human diseases or to test therapeutic or toxic agents. (See USPN 4,736,866; USPN 5,175,383; and USPN 5,767,337; incorporated herein by reference). In some cases, the introduced gene may be activated at a specific time in a specific tissue type during fetal development or postnatally.

25 Expression of the transgene is monitored by analysis of phenotype or tissue-specific mRNA expression, in transgenic animals before, during, and after being challenged with experimental drug therapies.

Embryonic Stem Cells

Embryonic stem cells (ES) isolated from rodent embryos retain the potential to form an embryo. When ES cells are placed inside a carrier embryo, they resume normal development and contribute to all tissues of the live-born animal. ES cells are the preferred cells used in the creation of experimental knockout and knockin rodent strains. Mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and are grown under culture conditions well known in the art. Vectors for knockout strains contain a disease gene candidate modified to include a marker gene which disrupts transcription and/or translation of the endogenous disease candidate gene in vivo. The vector is

introduced into ES cells by transformation methods such as electroporation, liposome delivery, microinjection, and the like which are well known in the art. The endogenous rodent gene is replaced by the disrupted disease gene through homologous recombination and integration during cell division. Expression of the marker gene confers a selective advantage to the transformed cells when incubated with an otherwise toxic/lethal selecting agent. Transformed ES cells are selected, identified, and preferably microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains.

ES cells are also used to study the differentiation of various cell types and tissues *in vitro*, such as neural cells, hematopoietic lineages, and cardiomyocytes (Bain et al. (1995) Dev. Biol. 168:342-357; Wiles and Keller (1991) Development 111:259-267; and Klug et al. (1996) J. Clin. Invest. 98:216-224). Recent developments demonstrate that ES cells derived from human blastocysts may also be manipulated *in vitro* to differentiate into eight separate cell lineages, including endoderm, mesoderm, and ectodermal cell types (Thomson et al. (1998) Science 282:1145-1147).

Knockout Analysis

In gene knockout analysis, a region of a human disease gene candidate is enzymatically modified to include a non-mammalian gene such as the neomycin phosphotransferase gene (*neo*; Capecchi (1989) Science 244:1288-1292). The inserted coding sequence disrupts transcription and translation of the targeted gene and prevents biochemical synthesis of the disease candidate protein. The modified gene is transformed into cultured embryonic stem cells (described above), the transformed cells are injected into rodent blastulae, and the blastulae are implanted into pseudopregnant dams. Transgenic progeny are crossbred to obtain homozygous inbred lines.

Knockin Analysis

Totipotent ES cells, present in the early stages of embryonic development, can be used to create knockin humanized animals (pigs) or transgenic animal models (mice or rats) of human diseases. With knockin technology, a region of a human gene is injected into animal ES cells, and the human sequence integrates into the animal cell genome by recombination. Totipotent ES cells which contain the integrated human gene are handled as described above. Inbred animals are studied and treated to obtain information on the analogous human condition. These methods have been used to model several human diseases. (See, e.g., Lee et al. (1998) Proc. Natl. Acad. Sci. 95:11371-11376; Baudoin et al. (1998) Genes Dev. 12:1202-1216; and Zhuang et al. (1998) Mol. Cell Biol. 18:3340-3349).

Non-Human Primate Model

The field of animal testing deals with data and methodology from basic sciences such as physiology, genetics, chemistry, pharmacology and statistics. These data are paramount in evaluating the

effects of therapeutic agents on non-human primates as they can be related to human health. Monkeys are used as human surrogates in vaccine and drug evaluations, and their responses are relevant to human exposures under similar conditions. Cynomolgus and Rhesus monkeys (Macaca fascicularis and Macaca mulatta, respectively) and Common Marmosets (Callithrix jacchus) are the most common non-human primates (NHPs) used in these investigations. Since great cost is associated with developing and maintaining a colony of NHPs, early research and toxicological studies are usually carried out in rodent models. In studies using behavioral measures such as drug addiction, NHPs are the first choice test animal. In addition, NHPs and individual humans exhibit differential sensitivities to many drugs and toxins and can be classified as a range of phenotypes from "extensive metabolizers" to "poor metabolizers" of these agents.

In additional embodiments, the nucleic acid molecules which encode the mammalian protein may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleic acid molecules that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

15

Examples

It is understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary. It is also understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. The examples below are provided to best describe the subject invention and its representative constituents.

I cDNA Library Construction

The RALINOT01 cDNA library was constructed from liver tissue removed from a pool of fifty 25 10- to 11-week-old Sprague-Dawley female rats (Pharmacon, Waverly PA). The animals were housed in standard laboratory caging and fed PMI-certified Rodent Diet #5002. The animals appeared to be in good health at the time tissue was harvested. The animals were anesthetized by CO₂ inhalation, and then cardiocentesis was performed.

Frozen tissue was homogenized and lysed in TRIZOL reagent (1 g tissue/10 ml TRIZOL; Life 30 Technologies), a monophasic solution of phenol and guanidine isothiocyanate, using a POLYTRON homogenizer (PT-3000; Brinkmann Instruments, Westbury NY). After a brief incubation on ice, chloroform (1:5 v/v) was mixed with the reagent, and then centrifuged at 1,000 rpm. The upper aqueous layer was removed to a fresh tube, and the RNA precipitated with isopropanol, resuspended in DEPC-treated water, and treated with DNase I for 25 min at 37°C. The RNA was re-extracted once with

phenol-chloroform, pH 4.7, and precipitated using 0.3 M sodium acetate and 2.5 volumes ethanol. The mRNA was then isolated using an OLIGOTEX kit (QIAGEN, Chatsworth CA) and used to construct the cDNA library.

The mRNA was handled according to the recommended protocols in the SUPERSCRIPT plasmid system (Life Technologies). The cDNAs were fractionated on a SEPHAROSE CL-4B column (Amersham Pharmacia Biotech), and those cDNAs exceeding 400 bp were ligated into the pINCY1 plasmid vector (Incyte Pharmaceuticals). The plasmid pINCY1 was subsequently transformed into DH5 α or DH10B competent cells (Life Technologies).

The RAKINOT01 library was constructed using mRNA isolated from kidney tissue removed from a pool of fifty, 7- to 8-week-old male Sprague-Dawley rats, as described above.

The RAKINOT02 library was constructed using mRNA isolated from kidney tissue removed from a pool of fifty, 10- to 11-week-old female Sprague-Dawley rats, as described above.

II cDNA Library Normalization

In some cases, cDNA libraries were normalized in a single round according to the procedure of Soares *et al.* (1994, Proc. Natl. Acad. Sci. 91:9228-9232) with the following modifications. The primer to template ratio in the primer extension reaction was increased from 2:1 to 10:1. Reduction of each dNTP concentration in the reaction to 150 μ M allowed the generation of longer (400-1000 nucleotide (nt)) primer extension products. The reannealing hybridization was extended from 13 to 19 hours. The single stranded DNA circles of the normalized library were purified by hydroxyapatite chromatography, converted to partially double-stranded by random priming, and electroporated into DH10B competent bacteria (Life Technologies).

The Soares normalization procedure is designed to reduce the initial variation in individual cDNA frequencies and to achieve abundances within one order of magnitude while maintaining the overall sequence complexity of the library. In the normalization process, the prevalence of high-abundance cDNA clones decreases significantly, clones with mid-level abundance are relatively unaffected, and clones for rare transcripts are increased in abundance. In the modified Soares normalization procedure, significantly longer hybridization times are used to increase gene discovery rates by biasing the normalized libraries toward low-abundance cDNAs that are well represented in a standard transcript image.

The RALINON03, RALINON04, and RALINON07 normalized rat liver cDNA libraries were constructed with 2.0×10^6 , 4.6×10^5 , and 2.0×10^6 independent clones from the RAKINOT01 cDNA library, respectively. The RAKINOT01 cDNA library was normalized in one round using conditions adapted from Soares (*supra*) except that a significantly longer (48-hour) reannealing hybridization was

used.

III cDNA Library Prehybridization

The RALINOH01 cDNA library was constructed with clones from the RALINOT01 cDNA library. After preparation of the RALINOT01 cDNA library, 9,984 clones were spotted onto a nylon filter, lysed, and the plasmid DNA was bound to the filter. The filter was incubated with pre-warmed hybridization buffer and then hybridized at 42°C for 14-16 hours in 0.75 M NaCl, 0.1 M Na₂HPO₄/NaH₂PO₄, 0.15 M tris-HCl (pH 7.5), 5x Denhardt's Solution, 2% SDS, 100 µg/ml sheared salmon sperm DNA, 50% formamide, and [³²P]-labeled oligonucleotide molecules made from reverse transcribed rat liver mRNA from an untreated animal. The filter was rinsed with 2 x SSC (saline sodium citrate) at ambient temperature for 5 minutes followed by washing for 30 minutes at 68°C with pre-warmed washing solution (2 x SSC, 1% SDS). The wash was repeated with fresh washing solution for an additional 30 minutes at 68°C. Filters were then washed twice with pre-warmed washing solution (0.6 x SSC, 1% SDS) for 30 minutes at 68°C. Some 4,224 clones had very low hybridization signals and about 20% of the clones had no signals and two groups were isolated and sequenced.

IV Isolation and Sequencing of cDNA Clones

DNA was isolated using the following protocol. Single bacterial colonies were transferred into individual wells of 384-well plates (Genetix Ltd, Christchurch, United Kingdom) using sterile toothpicks. The wells contained 1 ml of sterile Terrific Broth (Life Technologies) with 25 mg/l carbenicillin and 0.4% glycerol (v/v). The plates were covered and placed in an incubator (Thermodyne, Newtown Square PA) at 37°C for 8-10 hours. Plasmid DNA was released from the cells and amplified using direct link PCR (Rao, V.B. (1994) Anal. Biochem. 216:1-14) as follows. The direct link PCR solution included 30 ml of NUCLEIX PLUS PCR nucleotide mix (Amersham Pharmacia Biotech, Piscataway NJ) and 300 µl of Taq DNA polymerase (Amersham Pharmacia Biotech). Five microlitres of the PCR solution were added to each of the 384 wells using the MICROLAB 2200 system (Hamilton, Reno NV); plates were centrifuged at 1000 rpm for 20 seconds and refrigerated until use. A 384 pin tool (V&P Scientific Inc, San Diego CA) was used to transfer bacterial cells from the incubation plate into the plate containing the PCR solution where 0.1% Tween 20 caused the cells to undergo lysis and release the plasmid DNA. After lysis, the plates were centrifuged up to 500 rpm, covered with a cycle sealer, and cycled using a 384-well DNA ENGINE thermal cycler (MJ Research, Watertown MA) using the program dPCR30 with the following parameters: Step 1) 95 °C, 1 minute; Step 2) 94 °C, 30 seconds; Step 3) 55 °C, 30 seconds; Step 4) 72 °C, 2 minutes; Step 5) steps 2, 3, and 4 repeated 29 times; Step 6) 72 °C, 10 minutes; and Step 7) storage at 4 °C.

The concentration of DNA in each well was determined by dispensing 100 µl PICO GREEN quantitation reagent (0.25% (v/v), Molecular Probes, Eugene OR) dissolved in 1x TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the quantitation reagent. The plate was scanned in a Fluoroscan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantitate the concentration of DNA. Typical concentrations of each DNA sample were in the range of 100 to 500 ng/ml.

5 The cDNAs were prepared for sequencing using either a HYDRA microdispenser (Robbins Scientific, Sunnyvale CA) or MICROLAB 2200 system (Hamilton) in combination with the DNA
10 ENGINE thermal cyclers (MJ Research). The cDNAs were sequenced using the method of Sanger, F. and A.R. Coulson (J. Mol. Biol. (1975) 94:441-448) and the ABI 377 sequencing systems (PE Biosystems). Most of the isolates were sequenced according to standard ABI protocols using ABI kits (PE Biosystems). The solution volumes were used at 0.25x - 1.0x concentrations. Typically, 500 to 700 base pairs were sequenced in 3.5 to 4 hours. In the alternative, cDNAs may have been sequenced using
15 solutions and dyes from Amersham Pharmacia Biotech.

V Rat Liver and Kidney Gene Selection

As a first step, originator molecules from high throughput sequencing experiments were derived from clone inserts from RALINOT01, RAKINOT01, RAKINOT02, RALINOH01, RALINON03, RALINON04 and RALINON07. cDNA library clones were obtained. There were 18,140 rat liver molecules and 5,779 rat kidney molecules.

20 Additionally, 1,500 rat molecules derived from clone inserts of any of 113 rat cDNA libraries were selected based on their homology to genes coding for polypeptides implicated in toxicological responses including peroxisome-associated genes, lysosome-associated genes, apoptosis-associated genes, cytochrome P450 genes, detoxification genes such as sulfotransferases, glutathione S-transferases, and cysteine proteases, and the like.

25 Then, all the remaining molecules derived from all of the rat cDNA library clones were clustered based on the originator molecules described above. The clustering process involved identifying overlapping molecules that have a match quality indicated by a product score of 50 using BLAST. 30 6581 master clusters were identified.

After forming the clone clusters, a consensus sequence was generated based on the assembly of the clone molecules using PHRAP (Phil Green, University of Washington). The assembled molecules were then annotated by first screening the assembled molecules against GenBank using BLASTn and then by screening the assembled molecules against GenPept using FASTX. About two thirds of the

assembled molecules were annotated, about one third of the assembled molecules were not annotated. For example, for nucleic acid sequence analysis, the program BLASTN 1.4.9MP-WashU was used with default parameters; ctxfactor=2.00; E=10; MatID, 0; Matrix name, +5, 4. In another example, for amino acid sequence analysis, the program NCBI-BLASTX 2.0.4 was used with default parameters; matrix, BLOSUM62; gap penalties, existence 11, extension 1; frameshift window, decay constant 50, 0.1.

VI Substrate and Array Element/Probe Preparation

Clones nominated in the process described in Example V were used to generate array elements. Each array element was amplified from bacterial cells. PCR amplification used primers complementary to the vector sequences flanking the cDNA insert. Array elements were amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 µg. Amplified array elements were then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements were immobilized on polymer-coated glass slides. Glass microscope slides (Corning, Corning NY) cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides were etched in 4% hydrofluoric acid (VWR, West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma-Aldrich, St. Louis MO) in 95% ethanol. Coated slides were cured in a 110°C oven.

Array elements were applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522 and incorporated herein by reference. In brief, 1 µl of the array element DNA, at an average concentration of 0.5 µg/ml in 3 x SSC, was loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposited about 5 nl of the array element sample per slide. A total of 7404 array elements representing rat liver and kidney genes and a variety of control elements, including 14 synthetic control molecules, human genomic DNA, and yeast genomic DNA, were arrayed in four identical quadrants within a 1.8 cm² area of the glass substrate.

Microarrays were UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays were washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites were blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS; Tropix Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

VII Target Preparation

Male Sprague-Dawley rats (6-8 wk old) were dosed intraperitoneally with one of the following: clofibrate (CLO; Acros, Geel, Belgium) at 250 mg/kg body weight (bw); acetaminophen (APAP; Acros) at 1000 mg/kg bw; benzo(a)pyrene (B(a)P; Acros) at 10 mg/kg bw; or dimethylsulfoxide vehicle (DMSO;

Acros) at less than 2 ml/kg bw, and the animals were later euthanized by CO₂ inhalation. Animals were monitored daily for physical condition and body weight. Three animals per group were sacrificed approximately 12 hours, 24 hours, 3d (d), 7d, 14d, and 28d following the single dose. Prior to sacrifice a blood sample from each animal was drawn and assayed for serum alanine transferase (ALT) and serum aspartate aminotransferase (AST) levels using a diagnostic kit (Sigma-Aldrich). Observed gross pathology and liver weights were recorded at time of necropsy. Liver, kidney, brain, spleen and pancreas from each rat were harvested, flash frozen in liquid nitrogen, and stored at -80°C.

In the alternative, male Han-Wistar rats (8-9 wk old) were dosed by oral gavage with one of the following: fenofibrate (FEN; Sigma-Aldrich) at 250 mg/kg bw; carbon tetrachloride (CCL₄; Sigma-Aldrich) at 3160 mg/kg bw, hydrazine (HYDR; Sigma-Aldrich) at 120 mg/kg bw; α-naphthylisothiocyanate (ANIT; Sigma-Aldrich) at 200 mg/kg bw; 4-acetylaminofluorene (4-AFF; Lancaster Synthesis, Morecambe, Lancashire, UK) at 1000 mg/kg bw; corn oil vehicle, or sterile water vehicle, at 10 ml/kg bw. The animals were checked twice daily for clinical signs of distress. Blood was collected six days prior to the dose and at sacrifice. Three animals per group were sacrificed approximately six hours and 24 hours following the single dose. The animals were euthanized by exsanguination under isoflurane anaesthesia. Observed gross pathology and liver weights were recorded at time of necropsy. Livers from each rat were harvested, dissected into approximate 100 mg pieces, flash frozen in liquid nitrogen, and stored at -70°C.

For each target preparation, frozen liver was homogenized and lysed in TRIZOL reagent (Life Technologies, Gaithersburg MD) following the modifications for liver RNA isolation. Messenger RNA was isolated using an OLIGOTEX kit (QIAGEN) and labeled with either Cy3- or Cy5-labeled primers (Operon Technologies, Alameda CA) using the GEMBRIGHT labeling kit (Incyte Pharmaceuticals). Messenger RNA isolated from tissues of rats treated with clofibrate, acetaminophen, or benzo(a)pyrene was labeled with Cy5 and mRNA isolated from tissues of rats treated with DMSO was labeled with Cy3. Quantitative and differential expression pattern control cDNAs were added to each labeling reaction. Labeled cDNA was treated with 0.5 M sodium bicarbonate (pH 9.2) for 20 min at 85°C to degrade the RNA and purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech, Palo Alto CA). Cy3-labeled control sample and Cy5-labeled experimental sample were combined and precipitated in glycogen, sodium acetate, and ethanol.

Targets are also prepared from tissue needle biopsy samples. Samples are used to identify changes within the tissue following exposure to, for example, a toxic compound, a potential toxic compound, a compound with unknown metabolic responses, and a pharmacological compound.

VIII Hybridization

Hybridizations were carried out using the methods described by Shalon (*supra*).

IX Detection

The microscope used to detect the reporter-labeled hybridization complexes was equipped with an Innova 70 mixed gas 10 W laser (Coherent Lasers, Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3, and 632 nm for excitation of Cy5. The excitation laser light was focused on the array using a 20x microscope objective (Nikon, Melville NY). The slide containing the array was placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example was scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excited the two fluorophores sequentially. Emitted light was split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics, San Jose CA) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes were used to filter the signals. The emission maxima of the fluorophores used were 565 nm for Cy3 and 650 nm for Cy5. Each array was typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus was capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans was typically calibrated using the signal intensity generated by a cDNA control species added to the probe mix at a known concentration. A specific location on the array contained a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two probes from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration was done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube was digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Norwood MA) installed in an IBM-compatible PC computer. The digitized data were displayed as an image where the signal intensity was mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data was also analyzed quantitatively. Where two different fluorophores were excited and measured simultaneously, the data were first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid was superimposed over the fluorescence signal image such that the signal from each spot

was centered in each element of the grid. The fluorescence signal within each element was then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis was the GEMTOOLS gene expression analysis program (Incyte Pharmaceuticals).

5

X Results

The expression patterns of eight cytochrome P450 isozymes known to be induced in a toxicological response were monitored during the 28 day time course. The results using clofibrate, acetaminophen, and benzo(a)pyrene are shown in Table 1, Table 2, and Table 3, respectively. Each of the known genes was upregulated or downregulated greater than 2-fold at least once during the time course.

TABLE 1 Gene expression patterns (x-fold change) of known genes in clofibrate-treated rat liver

Gene	12 hours	24 hours	3 days	7 days	28 days
P450 LA-omega 4A3	14.8	26.6	1.1	0.5	0.47
P450 4A	7.0	16.6	1.4	0.5	1.3
P450 3A2	0.14	1.2	0.63	0.50	0.45

TABLE 2 Gene expression patterns (x-fold change) of known genes in acetaminophen-treated rat liver

Gene	12 hours	24 hours	3 days	7 days	14 days	28 days
P450 4A	1.0	4.5	2.1	2.0	4.4	4.8
P450f 2C7	0.21	0.43	0.47	0.5	1.2	1.3
P450 14DM	0.31	0.20	2.0	1.1	1.4	0.42

TABLE 3 Gene expression patterns (x-fold change) of known genes in benzo(a)pyrene-treated rat liver

Gene	12 hours	24 hours	3 days	7 days	14 days	28 days
P450 LA-omega 4A3	1.2	2.3	2.4	1.4	6.8	1.2
P450 MCA-inducible 1A2	7.3	9.2	5.7	2.5	2.5	0.5

In addition, results from two samples that had been treated identically were compared to determine the range of normal variation of gene expression patterns between the samples. In one analysis, where two different samples were prepared from identically treated tissues, gene expression patterns of cDNAs which were upregulated or downregulated not more than 1.7-fold were within the 95% confidence limits of a Poisson normal distribution. In a separate analysis, gene expression patterns of cDNAs which were upregulated or downregulated more than 2-fold accounted for not more than 5% of

the total hybridizable sample nucleic acid molecules in two identically-treated tissue samples.

We have discovered novel nucleotide molecules that are up-regulated or down-regulated at least 2-fold at least once during the time course. These molecules are SEQ ID NOs:1-16 provided in the Sequence Listing. These polynucleotide molecules can be used for screening compounds or therapeutics 5 for a toxicologic effect and applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

Table 4 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with clofibrate (CLO). Table 5 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during 10 the time course following treatment with CLO.

TABLE 4 Gene expression patterns (x-fold change) of CLO-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	28 days
2	2.6	1.4	0.5	1.1	1.2
3	1.3	2	1.3	1.5	1.5
4	2	0.36	0.47	0.26	0.30
5	1.7	2.9	1.6	1.5	1.2
8	2.6	1.7	1.3	1.3	1.4

20

TABLE 5 Gene expression patterns (x-fold change) of CLO-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	28 days
1	n.d.	0.26	0.45	0.26	1.1
4	2.0	0.36	0.47	0.26	0.30
7	0.24	0.42	0.37	1.1	1.5

(n.d. = not detected)

Table 6 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with acetaminophen (APAP). Table 7 30 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with APAP.

TABLE 6 Gene expression patterns (x-fold change) of APAP-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	14 days	28 days
5	2	1.3	2.2	1.1	0.5	1.2
	3	1.2	2.1	0.47	0.46	1.8
	4	3.3	0.47	0.47	0.23	0.35
	5	1.1	2.1	1.1	1.2	1.3
	6	1.8	5	2.5	1.1	1.4
	8	1.1	2.5	1.1	1	1.7

10 TABLE 7 Gene expression patterns (x-fold change) of APAP-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	14 days	28 days
1	0.36	0.19	0.46	0.25	0.5	1.4
4	3.3	0.48	0.47	0.23	0.35	0.36
7	0.33	0.21	1.7	n.d.	1	0.39

15 (n.d. = not detected)

Table 8 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with benzo(a)pyrene (B(a)P). Table 9 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with B(a)P.

20 TABLE 8 Gene expression patterns (x-fold change) of B(a)P-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	1 day	3 days	7 days	14 days	28 days
25	2	0.5	0.47	1.2	1.1	2.6
	3	1.4	2.1	1.2	1.5	2.7
	5	1.5	1.4	1.2	0.47	2
	6	2.2	1.4	1.4	1.2	2.2
	7	1.2	2.2	1.4	0.5	0.42
	8	1.6	1.7	1.3	1.3	2

30 (n.d. = not detected)

TABLE 9 Gene expression patterns (x-fold change) of B(a)P-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	1 day	3 days	7 days	14 days	28
1	0.37	0.39	0.35	1.4	0.33	1.5
4	0.5	0.26	0.31	0.36	0.47	n.d.

5 (n.d. = not detected)

Table 10 shows the library abundance of selected molecules that were up- or down-regulated at least once following treatment with various agents. Library abundance of each SEQ ID NO is presented as relative to that library which included the least abundant levels of nucleic acid molecule (SEQ ID NO) present.

10 TABLE 10 Library abundance (least abundant = 1) patterns of nucleic acid molecules

SEQ ID NO:	Untreated	CLO	FEN	APAP	BaP	CCl ₄	HYDR	ANIT	4-AAF
8	4	7	6	3	9	4	1	1	3
9	13	5	6	4	15	5	6	6	2
10	n.d.	1	8	3	n.d.	n.d.	n.d.	1	n.d.
11	5	2	4	8	20	7	10	n.d.	2

15 (n.d. = not detected)

20 **XI Identification and Analyses of Homologous Molecule in other Organisms**

The rat sequences (SEQ ID NOs:1-16) were used to identify additional sequences in the ZOOSEQ and LIFESEQ databases (Incyte Pharmaceuticals) related to rat nucleic acid molecules regulated during toxicological response (SEQ ID NOs:18-47).

25 The first pass cDNAs, SEQ ID NOs:5, and 60 through 134, were assembled using PHRAP (Phil Green, *supra*), using the following default parameters, to produce the contiguous sequence SEQ ID NO:135. Mismatch penalty = -2; gap initiation penalty <0; gap extension penalty <0; minimum length of matching word = 14; minimum SWAT score = 30; bandwidth = 14; use raw SW scores, "No"; index word size = 10; maximum gap size = 30; number of initial bases to be converted to 'N', 0; vector segment length = 60; Mismatch penalty for scoring degenerate end sequence = -2; Min. score for converting degenerate end sequence to 'N', 20; Minimum size of confirming segment = 8; Amount by which confirming segments are trimmed = 1; Penalty for confirming matches = -5; Min. SWAT score for confirming matches = 30; LLR cutoff for displaying discrepancies = 20; Minimum segment size = 8; Spacing between nodes = 4; Split/reassemble initial 'greedy' assembly, "No".

Translation of SEQ ID NO:135 using MACDNASIS PRO software (version 1.0, Hitachi Software Engineering) using default parameters of the program elucidated the putative protein coding region, SEQ ID NO:136. The nucleic acid and amino acid sequences were queried against databases such as the LIFESEQ (Incyte), GenBank, and SwissProt databases using BLAST. Motifs, HMM algorithms, and alignments with BLOCKS, PRINTS, Prosite, and PFAM databases were used to perform functional analyses; the antigenic index (Jameson-Wolf analysis) was determined using LASERGENE software (version 1.62d1, DNASTAR). BLAST2 analysis of SEQ ID NOs:135 and 136 using the human EST LIFESEQ database (Incyte) identified Incyte Clone Numbers 746355H1 (SEQ ID NO:137) and 1294663H1 (SEQ ID NO:138) which were assembled with their respective clustered clones to produce SEQ ID NOs:37 and 38 which encoded SEQ ID NOs:51 and 52, respectively.

Functional analysis of SEQ ID NO:136 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential protein kinase C phosphorylation site at residue S84 (Motifs); a potential signal peptide from residue M1 through residue A33 (SPScan); a potential transmembrane domain from residue P37 through residue L56 (HMM TM), a sodium/neurotransmitter symporter signature from residue G34 through A53, a sodium/alanine symporter signature from G34 through A53, and an asparaginase/glutaminase family signature from residue W64 through residue G75 (BLOCKS and PRINTS).

Functional analysis of SEQ ID NO:51 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential protein kinase C phosphorylation site at residue S83 (Motifs); a potential signal peptide sequence from residue M1 through residue A52 (SPScan); a sodium/alanine symporter signature from residue G33 through residue A52, an asparaginase/glutaminase family signature from residue W63 through residue G74, and a channel-forming colicin domain from residue K31 through residue G49 (BLOCKS and PRINTS). Functional analysis of SEQ ID NO:52 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential signal peptide sequence from residue M1 through A53 (SPScan); a sodium/alanine symporter signature from residue G34 through residue A53, a 6-phosphogluconate dehydrogenase family signature from residue G15 through residue A40, an FAD-dependant glycerol-3-phosphate dehydrogenase family signature from residue Y18 through residue Y30, and a vacuolar ATP synthetase 16 kDa subunit signature from residue L39 through residue G65 (BLOCKS and PRINTS).

CLAIMS

What is claimed is:

1. A method for detecting or diagnosing the effect of a toxic compound or molecule associated with increased or decreased levels of nucleic acid molecules in a mammalian subject comprising:

- 5 a) treating a mammalian subject with a toxic compound or molecule;
- b) obtaining a sample containing nucleic acids from the mammalian subject treated with the toxic compound or molecule;
- c) contacting the sample with a microarray comprising a plurality of nucleic acid molecules of SEQ ID NOs:1-47, or a fragment thereof under conditions for the formation of one or more hybridization complexes; and
- d) detecting the hybridization complexes, wherein the presence, absence or change in amount of the hybridization complex, as compared with the hybridization complexes formed from nucleic acid molecules from an untreated mammalian subject, is indicative of a metabolic response to the toxic compound or molecule.

15 2. The method of claim 1 wherein:

- a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;
- b) the sample is liver tissue;
- c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;
- d) the toxic compound is a peroxisome proliferator;
- e) the toxic compound is a hypolipidemic drug; and
- f) the toxic compound is clofibrate or one of its corresponding metabolites.

20 3. The method of claim 1 wherein:

- a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;
- b) the sample is liver tissue;
- c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;
- d) the toxic compound is acetaminophen or one of its corresponding metabolites.

25 4. The method of claim 1 wherein:

- a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;
- b) the sample is liver tissue;
- c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;
- d) the toxic compound is a polycyclic aromatic hydrocarbon;

- e) the toxic compound is a diol epoxide; and
- f) the toxic compound is benzo(a)pyrene, or one of its corresponding metabolites.

5. A method for detecting or diagnosing a toxicological response to a test compound or molecule in a mammalian subject, the method comprising:

- 5 a) treating a mammalian subject with a test compound or molecule;
- b) obtaining a sample containing nucleic acids from the mammalian subject treated with the test compound or molecule;
- c) contacting the sample with a microarray comprising a plurality of nucleic acid molecules of SEQ ID NOs:1-47, or a fragment thereof, under conditions for the formation of one or more hybridization complexes;
- d) detecting the hybridization complexes, wherein the presence, absence or change in amount of the hybridization complex, as compared with the hybridization complexes formed from nucleic acid molecules from a normal or untreated mammalian subject, is indicative of a toxic response to the test compound or molecule.

15. 6. The method of claim 5 wherein the test compound which elicits the metabolic response is a compound with a previously known metabolic response.

7. The method of claim 5 wherein the test compound which elicits the metabolic response is a compound with a previously unknown metabolic response.

8. An isolated and purified nucleic acid molecule selected from SEQ ID NOs:1-11, 17-33, 36, 39, 20 and 41, or a fragment thereof, wherein said fragments are at least 60 contiguous nucleotides in length.

9. A method of using a molecule selected from SEQ ID NOs:1-59 or a fragment thereof to screen a library of molecules or compounds to identify at least one molecule or compound which specifically binds the selected molecule, the method comprising:

- 25 a) combining the selected molecule with a library of molecules or compounds under conditions to allow specific binding; and
- b) detecting specific binding, thereby identifying a molecule or compound which specifically binds the selected molecule.

10. The method of claim 9 wherein the library is selected from DNA molecules, RNA molecules, peptide nucleic acids, artificial chromosome constructions, peptides, proteins, and drugs.

30. 11. An isolated and purified antibody identified using the method of claim 9.

12. An isolated and purified nucleic acid molecule variant having at least 70% nucleic acid sequence identity to the nucleic acid molecule of claim 8.

13. An isolated and purified nucleic acid molecule having a sequence which is complementary to the nucleic acid molecule of claim 8.

14. An isolated and purified agonist identified using the method of claim 9.
15. An isolated and purified antagonist identified using the method of claim 9.
16. An expression vector comprising at least a fragment of the nucleic acid molecule of claim 8.
17. A host cell comprising the expression vector of claim 16.
- 5 18. A method for producing a polypeptide, the method comprising the steps of:
 - a) culturing the host cell of claim 17 under conditions suitable for the expression of the polypeptide; and
 - b) recovering the polypeptide from the host cell culture.
19. An isolated and purified protein molecule encoded by the nucleic acid molecule selected from SEQ ID NOS:1-11, 17-33, 36, 39, and 41, an isolated and purified protein molecule of SEQ ID NOS:50 and 53, or a portion thereof, wherein said portions encode at least 20 contiguous amino acids in length.
- 10 20. A pharmaceutical composition comprising the protein molecule of claim 19 in conjunction with a suitable pharmaceutical carrier.

SEQUENCE LISTING

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CUNNINGHAM, Mary Jane
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PANZER, Scott
SEILHAMMER, Jeffrey J.
YUE, Henry
BAUGHN, Mariah
AZIMZAI, Yalda
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<212> DNA

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<213> Homo sapi

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<211> 2862
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<400> 36

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<220>

<221> misc_feature

<223> Incyte ID No.: 1968009

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<211> 978

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<213> Homo sapiens

<220>

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<223> Incyte ID No.: 1923127

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<223> Incyte ID No.: 375724.9

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 <213> Homo sapiens

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<400> 44

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 <211> 649
 <212> DNA
 <213> Homo sapiens

<220>
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 <223> Incyte ID No.: 2345712

<400> 45

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<213> Homo sapiens

<220>
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<400> 46

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<213> Homo sapiens

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<221> misc_feature
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<400> 47

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 <212> PRT
 <213> Homo sapiens

<220>
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 <223> Incyte ID No.: 2302721

<400> 48

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Ser	Val	Glu	Glu	Gly	Lys	Glu	Asn	Ile	Leu	His	Val	Ser	Glu	Asn
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Val	Ile	Phe	Thr	Asp	Val	Asn	Ser	Ile	Leu	Arg	Tyr	Leu	Ala	Arg
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Val	Ala	Thr	Thr	Ala	Gly	Leu	Tyr	Gly	Ser	Asn	Leu	Met	Glu	His
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Thr	Glu	Ile	Asp	His	Trp	Leu	Glu	Phe	Ser	Ala	Thr	Lys	Leu	Ser
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His	Ile	Gly	His	Ala	Lys	Ala	Ala	Leu	Leu	Asn	Gln	His	Tyr	Gln
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Pro	Glu	Lys	Glu	Lys	Glu	Asp	Phe	Glu	Lys	Val	Ile	Leu	Glu	Asp
					245				250					255
Val	Ala	Met	Leu	His	Ile	Lys	Pro	Asp	Gln	Phe	Thr	Tyr	Thr	Ser
					260				265					270
Asp	His	Phe	Glu	Thr	Ile	Met	Lys	Tyr	Ala	Glu	Lys	Leu	Ile	Gln
					275				280					285
Glu	Gly	Lys	Ala	Tyr	Val	Asp	Asp	Thr	Pro	Ala	Glu	Gln	Met	Lys
					290				295					300
Ala	Glu	Arg	Glu	Gln	Arg	Ile	Glu	Ser	Lys	His	Arg	Lys	Asn	Pro

	305	310	315
Ile Glu Lys Asn	Leu Gln Met Trp Glu	Glu Met Lys Lys Gly	Ser
320	325	330	
Gln Phe Gly Gln	Ser Cys Cys Leu Arg	Ala Lys Ile Asp Met	Ser
335	340	345	
Ser Asn Asn Gly	Cys Met Arg Asp Pro	Thr Leu Tyr Arg Cys	Lys
350	355	360	
Ile Gln Pro His	Pro Arg Thr Gly Asn	Lys Tyr Asn Val Tyr	Pro
365	370	375	
Thr Tyr Asp Phe	Ala Cys Pro Ile Val	Asp Ser Ile Glu Gly	Val
380	385	390	
Thr His Ala Leu	Arg Thr Thr Glu Tyr	His Asp Arg Asp Glu	Gln
395	400	405	
Phe Tyr Trp Ile	Ile Glu Ala Leu Gly	Ile Arg Lys Pro Tyr	Ile
410	415	420	
Trp Glu Tyr Ser	Arg Leu Asn Leu Asn	Asn Thr Val Leu Ser	Lys
425	430	435	
Arg Lys Leu Thr	Trp Phe Val Asn Glu	Gly Leu Val Asp Gly	Trp
440	445	450	
Asp Asp Pro Arg	Phe Pro Thr Val Arg	Gly Val Leu Arg Arg	Gly
455	460	465	
Met Thr Val Glu	Gly Leu Lys Gln Phe	Ile Ala Ala Gln Gly	Ser
470	475	480	
Ser Arg Ser Val Val	Asn Met Glu Trp	Asp Lys Ile Trp Ala	Phe
485	490	495	
Asn Lys Lys Val	Ile Asp Pro Val Ala	Pro Arg Tyr Val Ala	Leu
500	505	510	
Leu Lys Lys Glu Val	Ile Pro Val Asn	Val Pro Glu Ala Gln	Glu
515	520	525	
Glu Met Lys Glu Val	Ala Lys His Pro	Lys Asn Pro Glu Val	Gly
530	535	540	
Leu Lys Pro Val Trp	Tyr Ser Pro Lys	Val Phe Ile Glu Gly	Ala
545	550	555	
Asp Ala Glu Thr	Phe Ser Glu Gly Glu	Met Val Thr Phe Ile	Asn
560	565	570	
Trp Gly Asn Leu	Asn Ile Thr Lys Ile	His Lys Asn Ala Asp	Gly
575	580	585	
Lys Ile Ile Ser	Leu Asp Ala Lys Leu	Asn Leu Glu Asn Lys	Asp
590	595	600	
Tyr Lys Lys Thr	Thr Lys Val Thr Trp	Leu Ala Glu Thr Thr	His
605	610	615	
Ala Leu Pro Ile	Pro Val Ile Cys Val	Thr Tyr Glu His Leu	Ile
620	625	630	
Thr Lys Pro Val	Leu Gly Lys Asp Glu	Asp Phe Lys Gln Tyr	Val
635	640	645	
Asn Lys Asn Ser	Lys His Glu Glu Leu	Met Leu Gly Asp Pro	Cys
650	655	660	
Leu Lys Asp Leu	Lys Lys Gly Asp Ile	Ile Gln Leu Gln Arg	Arg
665	670	675	
Gly Phe Phe Ile	Cys Asp Gln Pro Tyr	Glu Pro Val Ser Pro	Tyr
680	685	690	
Ser Cys Lys Glu	Ala Pro Cys Val Leu	Ile Tyr Ile Pro Asp	Gly
695	700	705	
His Thr Lys Glu	Met Pro Thr Ser Gly	Ser Lys Glu Lys Thr	Lys
710	715	720	
Val Glu Ala Thr	Lys Asn Glu Thr Ser	Ala Pro Phe Lys Glu	Arg
725	730	735	
Pro Thr Pro Ser	Leu Asn Asn Asn Cys	Thr Thr Ser Glu Asp	Ser
740	745	750	
Leu Val Leu Tyr	Asn Arg Val Ala Val	Gln Gly Asp Val Val	Arg
755	760	765	
Glu Leu Lys Ala	Lys Lys Ala Pro Lys	Glu Asp Val Asp Ala	Ala
770	775	780	
Val Lys Gln Leu	Leu Ser Leu Lys Ala	Glu Tyr Lys Glu Lys	Thr
785	790	795	
Gly Gln Glu Tyr	Lys Pro Gly Asn Pro	Pro Ala Glu Ile Gly	Gln
800	805	810	
Asn Ile Ser Ser	Asn Ser Ser Ala Ser	Ile Leu Glu Ser Lys	Ser

	815		820		825									
Leu	Tyr	Asp	Glu	Val	Ala	Ala	Gln	Gly	Glu	Val	Val	Arg	Lys	Leu
					830					835				840
Lys	Ala	Glu	Lys	Ser	Pro	Lys	Ala	Lys	Ile	Asn	Glu	Ala	Val	Glu
					845					850				855
Cys	Leu	Leu	Ser	Leu	Lys	Ala	Gln	Tyr	Lys	Glu	Lys	Thr	Gly	Lys
					860					865				870
Glu	Tyr	Ile	Pro	Gly	Gln	Pro	Pro	Leu	Ser	Gln	Ser	Ser	Asp	Ser
					875					880				885
Ser	Pro	Thr	Arg	Asn	Ser	Glu	Pro	Ala	Gly	Leu	Glu	Thr	Pro	Glu
					890					895				900
Ala	Lys	Val	Leu	Phe	Asp	Lys	Val	Ala	Ser	Gln	Gly	Glu	Val	Val
					905					910				915
Arg	Lys	Leu	Lys	Thr	Glu	Lys	Ala	Pro	Lys	Asp	Gln	Val	Asp	Ile
					920					925				930
Ala	Val	Gln	Glu	Leu	Leu	Gln	Leu	Lys	Ala	Gln	Tyr	Lys	Ser	Leu
					935					940				945
Ile	Gly	Val	Glu	Tyr	Lys	Pro	Val	Ser	Ala	Thr	Gly	Ala	Glu	Asp
					950					955				960
Lys	Asp	Lys	Lys	Lys	Lys	Glu	Lys	Glu	Asn	Lys	Ser	Glu	Lys	Gln
					965					970				975
Asn	Lys	Pro	Gln	Lys	Gln	Asn	Asp	Gly	Gln	Arg	Lys	Asp	Pro	Ser
					980					985				990
Lys	Asn	Gln	Gly	Gly	Gly	Leu	Ser	Ser	Ser	Gly	Ala	Gly	Glu	Gly
					995					1000				1005
Gln	Gly	Pro	Lys	Lys	Gln	Thr	Arg	Leu	Gly	Leu	Glu	Ala	Lys	Lys
					1010					1015				1020
Glu	Glu	Asn	Leu	Ala	Asp	Trp	Tyr	Ser	Gln	Val	Ile	Thr	Lys	Ser
					1025					1030				1035
Glu	Met	Ile	Glu	Tyr	His	Asp	Ile	Ser	Gly	Cys	Tyr	Ile	Leu	Arg
					1040					1045				1050
Pro	Trp	Ala	Tyr	Ala	Ile	Trp	Glu	Ala	Ile	Lys	Asp	Phe	Phe	Asp
					1055					1060				1065
Ala	Glu	Ile	Lys	Lys	Leu	Gly	Val	Glu	Asn	Cys	Tyr	Phe	Pro	Met
					1070					1075				1080
Phe	Val	Ser	Gln	Ser	Ala	Leu	Glu	Lys	Glu	Lys	Thr	His	Val	Ala
					1085					1090				1095
Asp	Phe	Ala	Pro	Glu	Val	Ala	Trp	Val	Thr	Arg	Ser	Gly	Lys	Thr
					1100					1105				1110
Glu	Leu	Ala	Glu	Pro	Ile	Ala	Ile	Arg	Pro	Thr	Ser	Glu	Thr	Val
					1115					1120				1125
Met	Tyr	Pro	Ala	Tyr	Ala	Lys	Trp	Val	Gln	Ser	His	Arg	Asp	Leu
					1130					1135				1140
Pro	Ile	Lys	Leu	Asn	Gln	Trp	Cys	Asn	Val	Val	Arg	Trp	Glu	Phe
					1145					1150				1155
Lys	His	Pro	Gln	Pro	Phe	Leu	Arg	Thr	Arg	Glu	Phe	Leu	Trp	Gln
					1160					1165				1170
Glu	Gly	His	Ser	Ala	Phe	Ala	Thr	Met	Glu	Glu	Ala	Ala	Glu	Glu
					1175					1180				1185
Val	Leu	Gln	Ile	Leu	Asp	Leu	Tyr	Ala	Gln	Val	Tyr	Glu	Glu	Leu
					1190					1195				1200
Leu	Ala	Ile	Pro	Val	Val	Lys	Gly	Arg	Lys	Thr	Glu	Lys	Glu	Lys
					1205					1210				1215
Phe	Ala	Gly	Gly	Asp	Tyr	Thr	Thr	Thr	Ile	Glu	Ala	Phe	Ile	Ser
					1220					1225				1230
Ala	Ser	Gly	Arg	Ala	Ile	Gln	Gly	Gly	Thr	Ser	His	His	Leu	Gly
					1235					1240				1245
Gln	Asn	Phe	Ser	Lys	Met	Phe	Glu	Ile	Val	Phe	Glu	Asp	Pro	Lys
					1250					1255				1260
Ile	Pro	Gly	Glu	Lys	Gln	Phe	Ala	Tyr	Gln	Asn	Ser	Trp	Gly	Leu
					1265					1270				1275
Thr	Thr	Arg	Thr	Ile	Gly	Val	Met	Thr	Met	Val	His	Gly	Asp	Asn
					1280					1285				1290
Met	Gly	Leu	Val	Leu	Pro	Pro	Arg	Val	Ala	Cys	Val	Gln	Val	Val
					1295					1300				1305
Ile	Ile	Pro	Cys	Gly	Ile	Thr	Asn	Ala	Leu	Ser	Glu	Glu	Asp	Lys
					1310					1315				1320
Glu	Ala	Leu	Ile	Ala	Lys	Cys	Asn	Asp	Tyr	Arg	Arg	Arg	Leu	Leu

1325	1330	1335
Ser Val Asn Ile Arg Val Arg Ala Asp Leu Arg Asp Asn Tyr Ser		
1340	1345	1350
Pro Gly Trp Lys Phe Asn His Trp Glu Leu Lys Gly Val Pro Ile		
1355	1360	1365
Arg Leu Glu Val Gly Pro Arg Asp Met Lys Ser Cys Gln Phe Val		
1370	1375	1380
Ala Val Arg Arg Asp Thr Gly Glu Lys Leu Thr Val Ala Glu Asn		
1385	1390	1395
Glu Ala Glu Thr Lys Leu Gln Ala Ile Leu Glu Asp Ile Gln Val		
1400	1405	1410
Thr Leu Phe Thr Arg Ala Ser Glu Asp Leu Lys Thr His Met Val		
1415	1420	1425
Val Ala Asn Thr Met Glu Asp Phe Gln Lys Ile Leu Asp Ser Gly		
1430	1435	1440
Lys Ile Val Gln Ile Pro Phe Cys Gly Glu Ile Asp Cys Glu Asp		
1445	1450	1455
Trp Ile Lys Lys Thr Thr Ala Arg Asp Gln Asp Leu Glu Pro Gly		
1460	1465	1470
Ala Pro Ser Met Gly Ala Lys Ser Leu Cys Ile Pro Phe Lys Pro		
1475	1480	1485
Leu Cys Glu Leu Gln Pro Gly Ala Lys Cys Val Cys Gly Lys Asn		
1490	1495	1500
Pro Ala Lys Tyr Tyr Thr Leu Phe Gly Arg Ser Tyr		
1505	1510	

<210> 49
<211> 238
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 2742442

<400> 49

Met Ala Ala Arg Thr Gly His Thr Ala Leu Arg Arg Val Val Ser			
1	5	10	15
Gly Cys Arg Pro Lys Ser Ala Thr Ala Ala Gly Ala Gln Ala Pro			
20		25	30
Val Arg Asn Gly Arg Tyr Leu Ala Ser Cys Gly Ile Leu Met Ser			
35		40	45
Arg Thr Leu Pro Leu His Thr Ser Ile Leu Pro Lys Glu Ile Cys			
50		55	60
Ala Arg Thr Phe Phe Lys Ile Thr Ala Pro Leu Ile Asn Lys Arg			
65		70	75
Lys Glu Tyr Ser Glu Arg Arg Ile Leu Gly Tyr Ser Met Gln Glu			
80		85	90
Met Tyr Asp Val Val Ser Gly Val Glu Asp Tyr Lys His Phe Val			
95		100	105
Pro Trp Cys Lys Lys Ser Asp Val Ile Ser Lys Arg Ser Gly Tyr			
110		115	120
Cys Lys Thr Arg Leu Glu Ile Gly Phe Pro Pro Val Leu Glu Arg			
125		130	135
Tyr Thr Ser Val Val Thr Leu Val Lys Pro His Leu Val Lys Ala			
140		145	150
Ser Cys Thr Asp Gly Arg Leu Phe Asn His Leu Glu Thr Ile Trp			
155		160	165
Cys Phe Ser Pro Gly Leu Pro Gly Tyr Pro Arg Thr Cys Thr Leu			
170		175	180
Asp Phe Ser Ile Ser Phe Glu Phe Arg Ser Leu Leu His Ser Gln			
185		190	195
Leu Ala Thr Leu Phe Phe Asp Glu Val Val Lys Gln Met Val Ala			
200		205	210

Ala Phe Glu Arg Arg Ala Cys Lys Leu Tyr Gly Pro Glu Thr Asn
 215 220 225
 Ile Pro Arg Glu Leu Met Leu His Glu Val His His Thr
 230 235

<210> 50
 <211> 653
 <212> PRT
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 3511087

<400> 50

Met	Pro	Phe	Ser	Ala	Ser	Leu	Leu	Gly	Thr	Leu	Pro	Ile	Gly	Ala
1				5					10					15
Arg	Tyr	Ala	Pro	Pro	Pro	Ser	Phe	Ser	Glu	Phe	Tyr	Pro	Pro	Leu
				20					25					30
Thr	Ser	Ser	Leu	Glu	Asp	Phe	Cys	Ser	Ser	Leu	Asn	Ser	Phe	Ser
				35					40					45
Met	Ser	Glu	Ser	Lys	Arg	Asp	Leu	Ser	Thr	Ser	Thr	Ser	Arg	Glu
				50					55					60
Gly	Thr	Pro	Leu	Asn	Asn	Ser	Asn	Ser	Ser	Leu	Leu	Leu	Met	Asn
				65					70					75
Gly	Pro	Gly	Ser	Leu	Phe	Ala	Ser	Glu	Asn	Phe	Leu	Gly	Ile	Ser
				80					85					90
Ser	Gln	Pro	Arg	Asn	Asp	Phe	Gly	Asn	Phe	Phe	Gly	Ser	Ala	Val
				95					100					105
Thr	Lys	Pro	Ser	Ser	Ser	Val	Thr	Pro	Arg	His	Pro	Leu	Glu	Gly
				110					115					120
Thr	His	Glu	Leu	Arg	Gln	Ala	Cys	Gln	Ile	Cys	Phe	Val	Lys	Ser
				125					130					135
Gly	Pro	Lys	Leu	Met	Asp	Phe	Thr	Tyr	His	Ala	Asn	Ile	Asp	His
				140					145					150
Lys	Cys	Lys	Lys	Asp	Ile	Leu	Ile	Gly	Arg	Ile	Lys	Asn	Val	Glu
				155					160					165
Asp	Lys	Ser	Trp	Lys	Lys	Ile	Arg	Pro	Arg	Pro	Thr	Lys	Thr	Asn
				170					175					180
Tyr	Glu	Gly	Pro	Tyr	Tyr	Ile	Cys	Lys	Asp	Val	Ala	Ala	Glu	Glu
				185					190					195
Glu	Cys	Arg	Tyr	Ser	Gly	His	Cys	Thr	Phe	Ala	Tyr	Cys	Gln	Glu
				200					205					210
Glu	Ile	Asp	Val	Trp	Trp	Thr	Leu	Glu	Arg	Lys	Gly	Ala	Phe	Ser
				215					220					225
Glu	Ala	Phe	Phe	Gly	Gly	Asn	Gly	Lys	Ile	Asn	Leu	Thr	Val	Phe
				230					235					240
Lys	Leu	Leu	Gln	Glu	His	Leu	Gly	Glu	Phe	Ile	Phe	Leu	Cys	Glu
				245					250					255
Lys	Cys	Phe	Asp	His	Lys	Pro	Arg	Met	Ile	Ser	Lys	Arg	Asn	Lys
				260					265					270
Asp	Asn	Ser	Thr	Ala	Cys	Ser	His	Pro	Val	Thr	Lys	His	Glu	Phe
				275					280					285
Glu	Asp	Asn	Lys	Cys	Leu	Val	His	Ile	Leu	Arg	Glu	Thr	Thr	Val
				290					295					300
Lys	Tyr	Ser	Lys	Ile	Arg	Ser	Phe	His	Gly	Gln	Cys	Gln	Leu	Asp
				305					310					315
Leu	Cys	Arg	His	Glu	Val	Arg	Tyr	Gly	Cys	Leu	Arg	Glu	Asp	Glu
				320					325					330
Cys	Phe	Tyr	Ala	His	Ser	Leu	Val	Glu	Leu	Lys	Val	Trp	Ile	Met
				335					340					345
Gln	Asn	Glu	Thr	Gly	Ile	Ser	His	Asp	Ala	Ile	Ala	Gln	Glu	Ser
				350					355					360
Lys	Arg	Tyr	Trp	Gln	Asn	Leu	Glu	Ala	Asn	Val	Pro	Gly	Ala	Gln
				365					370					375
Val	Leu	Gly	Asn	Gln	Ile	Met	Pro	Gly	Phe	Leu	Asn	Met	Lys	Ile

<210> 51
<211> 112
<212> PRT
<213> *Homo sapiens*

<220>
<221> misc_feature
<223> Incyte ID No.: 1968009

<400> 51

<210> 52
<211> 114
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1923127

<400> 52

Met Glu Lys Pro Leu Phe Pro Leu Val Pro Leu His Trp Phe Gly
1 5 10 15
Phe Gly Tyr Thr Ala Leu Val Val Ser Gly Gly Ile Val Gly Tyr
20 25 30
Val Lys Thr Gly Ser Val Pro Ser Leu Ala Ala Gly Leu Leu Phe
35 40 45
Gly Ser Leu Ala Gly Leu Gly Ala Tyr Gln Leu Tyr Gln Asp Pro
50 55 60
Arg Asn Val Trp Gly Phe Leu Ala Ala Thr Ser Val Thr Phe Val
65 70 75
Gly Val Met Gly Met Arg Ser Tyr Tyr Gly Lys Phe Met Pro
80 85 90
Val Gly Leu Ile Ala Gly Ala Ser Leu Leu Met Ala Ala Lys Val
95 100 105
Gly Val Arg Met Leu Met Thr Ser Asp
110

<210> 53
<211> 114
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 3123954

<400> 53

Met Ala Ala Ile Pro Ser Ser Gly Ser Leu Val Ala Thr His Asp
1 5 10 15
Tyr Tyr Arg Arg Arg Leu Gly Ser Thr Ser Ser Asn Ser Ser Cys
20 25 30
Ser Ser Thr Glu Cys Pro Gly Glu Ala Ile Pro His Pro Pro Gly
35 40 45
Leu Pro Lys Ala Asp Pro Gly His Trp Trp Ala Ser Phe Phe Phe
50 55 60
Gly Lys Ser Thr Leu Pro Phe Met Ala Thr Val Leu Glu Ser Ala
65 70 75
Glu His Ser Glu Pro Pro Gln Ala Ser Ser Ser Met Thr Ala Cys
80 85 90
Gly Leu Ala Arg Asp Ala Pro Arg Lys Gln Pro Gly Gly Gln Ser
95 100 105
Ser Thr Ala Ser Ala Gly Pro Pro Ser
110

<210> 54
<211> 291
<212> PRT
<213> Homo sapiens

<220>
<221>
<223>

<400> 54

Met	Ser	Gln	Glu	Gly	Val	Glu	Leu	Glu	Lys	Ser	Val	Arg	Gly	Leu
1				5					10					15
Arg	Glu	Lys	Phe	His	Gly	Lys	Val	Ser	Ser	Lys	Lys	Ala	Gly	Ala
				20					25					30
Leu	Met	Arg	Lys	Phe	Gly	Ser	Asp	His	Thr	Gly	Val	Gly	Arg	Ser
				35					40					45
Ile	Val	Tyr	Gly	Val	Lys	Gln	Lys	Asp	Gly	Gln	Glu	Leu	Ser	Asn
				50					55					60
Asp	Leu	Asp	Ala	Gln	Asp	Pro	Pro	Glu	Asp	Met	Lys	Gln	Asp	Arg
				65					70					75
Asp	Ile	Gln	Ala	Val	Ala	Thr	Ser	Leu	Leu	Pro	Leu	Thr	Glu	Ala
				80					85					90
Asn	Leu	Arg	Met	Phe	Gln	Arg	Ala	Gln	Asp	Asp	Leu	Ile	Pro	Ala
				95					100					105
Val	Asp	Arg	Gln	Phe	Ala	Cys	Ser	Ser	Cys	Asp	His	Val	Trp	Trp
				110					115					120
Arg	Arg	Val	Pro	Gln	Arg	Lys	Glu	Val	Ser	Arg	Cys	Arg	Lys	Cys
				125					130					135
Arg	Lys	Arg	Tyr	Glu	Pro	Val	Pro	Ala	Asp	Lys	Met	Trp	Gly	Leu
				140					145					150
Ala	Glu	Phe	His	Cys	Pro	Lys	Cys	Arg	His	Asn	Phe	Arg	Gly	Trp
				155					160					165
Ala	Gln	Met	Gly	Ser	Pro	Ser	Pro	Cys	Tyr	Gly	Cys	Gly	Phe	Pro
				170					175					180
Val	Tyr	Pro	Thr	Arg	Ile	Leu	Pro	Pro	Arg	Trp	Asp	Arg	Asp	Pro
				185					190					195
Asp	Arg	Arg	Ser	Thr	His	Thr	His	Ser	Cys	Ser	Ala	Ala	Asp	Cys
				200					205					210
Tyr	Asn	Arg	Arg	Glu	Pro	His	Val	Pro	Gly	Thr	Ser	Cys	Ala	His
				215					220					225
Pro	Lys	Ser	Arg	Lys	Gln	Asn	His	Leu	Pro	Lys	Val	Leu	His	Pro
				230					235					240
Ser	Asn	Pro	His	Ile	Ser	Ser	Gly	Ser	Thr	Val	Ala	Thr	Cys	Leu
				245					250					255
Ser	Gln	Gly	Gly	Leu	Leu	Glu	Asp	Leu	Asp	Asn	Leu	Ile	Leu	Glu
				260					265					270
Asp	Leu	Lys	Glu	Val	Glu	Asp	Glu							
				275					280					285
Glu	Gly	Gly	Pro	Arg	Glu									
				290										

<210> 55

<211> 610

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 1867333

<400> 55

Met	Trp	Leu	Pro	Leu	Val	Leu	Leu	Ala	Val	Leu	Leu	Leu	Ala	
1				5					10				15	
Val	Leu	Cys	Lys	Val	Tyr	Leu	Gly	Leu	Phe	Ser	Gly	Ser	Ser	Pro
				20					25					30
Asn	Pro	Phe	Ser	Glu	Asp	Val	Lys	Arg	Pro	Pro	Ala	Pro	Leu	Val
				35					40					45
Thr	Asp	Lys	Glu	Ala	Arg	Lys	Lys	Val	Leu	Lys	Gln	Ala	Phe	Ser
				50					55					60
Ala	Asn	Gln	Val	Pro	Glu	Lys	Leu	Asp	Val	Val	Val	Ile	Gly	Ser
				65					70					75
Gly	Phe	Gly	Gly	Leu	Ala	Ala	Ala	Ala	Ile	Leu	Ala	Lys	Ala	Gly
				80					85					90
Lys	Arg	Val	Leu	Val	Leu	Glu	Gln	His	Thr	Lys	Ala	Gly	Gly	Cys

	95		100		105
Cys His Thr Phe Gly		Lys Asn Gly Leu	Glu Phe Asp Thr Gly	Ile	
110		115		120	
His Tyr Ile Gly Arg		Met Glu Glu Gly	Ser Ile Gly Arg Phe	Ile	
125		130		135	
Leu Asp Gln Ile Thr		Glu Gly Gln Leu	Asp Trp Ala Pro Leu	Ser	
140		145		150	
Ser Pro Phe Asp Ile		Met Val Leu Glu	Gly Pro Asn Gly Arg	Lys	
155		160		165	
Glu Tyr Pro Met Tyr		Ser Gly Glu Lys	Ala Tyr Ile Gln Gly	Leu	
170		175		180	
Lys Glu Lys Phe Pro		Gln Glu Glu Ala	Ile Ile Asp Lys Tyr	Ile	
185		190		195	
Lys Leu Val Lys Val		Val Ser Ser Gly	Ala Pro His Ala Ile	Leu	
200		205		210	
Leu Lys Phe Leu Pro		Leu Pro Val Val	Gln Leu Leu Asp Arg	Cys	
215		220		225	
Gly Leu Leu Thr Arg		Phe Ser Pro Phe	Leu Gln Ala Ser Thr	Gln	
230		235		240	
Ser Leu Ala Glu Val		Leu Gln Gln Leu	Gly Ala Ser Ser Glu	Leu	
245		250		255	
Gln Ala Val Leu Ser		Tyr Ile Phe Pro	Thr Tyr Gly Val Thr	Pro	
260		265		270	
Asn His Ser Ala Phe		Ser Met His Ala	Leu Leu Val Asn His	Tyr	
275		280		285	
Met Lys Gly Gly Phe		Tyr Pro Arg Gly	Gly Ser Ser Glu Ile	Ala	
290		295		300	
Phe His Thr Ile Pro Val		Ile Gln Arg	Ala Gly Gly Ala Val	Leu	
305		310		315	
Thr Lys Ala Thr Val		Gln Ser Val Leu	Leu Asp Ser Ala Gly	Lys	
320		325		330	
Ala Cys Gly Val Ser		Val Lys Lys Gly	His Glu Leu Val Asn	Ile	
335		340		345	
Tyr Cys Pro Ile Val Val		Ser Asn Ala	Gly Leu Phe Asn Thr	Tyr	
350		355		360	
Glu His Leu Leu Pro Gly		Asn Ala Arg	Cys Leu Pro Gly Val	Lys	
365		370		375	
Gln Gln Leu Gly Thr Val		Arg Pro Gly	Leu Gly Met Thr Ser	Val	
380		385		390	
Phe Ile Cys Leu Arg Gly		Thr Lys Glu Asp	Leu His Leu Pro	Ser	
395		400		405	
Thr Asn Tyr Tyr Val Tyr		Tyr Asp Thr Asp	Met Asp Gln Ala	Met	
410		415		420	
Glu Arg Tyr Val Ser		Met Pro Arg Glu	Glu Ala Ala Glu His	Ile	
425		430		435	
Pro Leu Leu Phe Phe Ala		Phe Pro Ser Ala	Lys Asp Pro Thr	Trp	
440		445		450	
Glu Asp Arg Phe Pro Gly		Arg Ser Thr	Met Ile Met Leu Ile	Pro	
455		460		465	
Thr Ala Tyr Glu Trp Phe		Glu Trp Gln Ala Glu Leu Lys	Gly		
470		475		480	
Lys Arg Gly Ser Asp Tyr		Glu Thr Phe Lys	Asn Ser Phe Val	Glu	
485		490		495	
Ala Ser Met Ser Val Val		Leu Lys Leu Phe	Pro Gln Leu Glu	Gly	
500		505		510	
Lys Val Glu Ser Val Thr		Ala Gly Ser Pro	Leu Thr Asn Gln	Phe	
515		520		525	
Tyr Leu Ala Ala Pro Arg		Gly Ala Cys	Tyr Gly Ala Asp His	Asp	
530		535		540	
Leu Gly Arg Leu His Pro		Cys Val Met Ala	Ser Leu Arg Ala	Gln	
545		550		555	
Ser Pro Ile Pro Asn Leu		Tyr Leu Thr Gly	Gln Asp Ile Phe	Thr	
560		565		570	
Cys Gly Leu Val Gly Ala		Leu Gln Gly	Ala Leu Leu Cys Ser	Ser	
575		580		585	
Ala Ile Leu Lys Arg Asn		Leu Tyr Ser Asp	Leu Lys Asn Leu	Asp	
590		595		600	
Ser Arg Ile Arg Ala Gln		Lys Lys Asn			

605

610

<210> 56
<211> 352
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1461451

<400> 56

Pro	Arg	Val	Arg	Gly	Arg	Trp	Val	Ala	His	Ala	Ser	Ala	His	Ala
1				5					10					15
Ser	Ala	His	Ala	Ser	Asp	Glu	Ile	Pro	Ala	Ser	Gly	Trp	Thr	Gln
					20				25					30
Trp	Phe	Cys	Thr	Glu	Ala	Leu	Val	Met	Val	Ala	Pro	Val	Trp	Tyr
				35					40					45
Leu	Val	Ala	Ala	Ala	Leu	Leu	Val	Gly	Phe	Ile	Leu	Phe	Leu	Thr
					50				55					60
Arg	Ser	Arg	Gly	Arg	Ala	Ala	Ser	Ala	Gly	Gln	Glu	Pro	Leu	His
				65					70					75
Asn	Glu	Glu	Leu	Ala	Gly	Ala	Gly	Arg	Val	Ala	Gln	Pro	Gly	Pro
				80					85					90
Leu	Glu	Pro	Glu	Glu	Pro	Arg	Ala	Gly	Gly	Arg	Pro	Arg	Arg	Arg
				95					100					105
Arg	Asp	Leu	Gly	Ser	Arg	Leu	Gln	Ala	Gln	Arg	Arg	Ala	Gln	Arg
				110					115					120
Val	Ala	Trp	Ala	Glu	Ala	Asp	Glu	Asn	Glu	Glu	Glu	Ala	Val	Ile
				125					130					135
Leu	Ala	Gln	Glu	Glu	Gly	Val	Glu	Lys	Pro	Ala	Glu	Thr	His	
				140					145					150
Leu	Ser	Gly	Lys	Ile	Gly	Ala	Lys	Lys	Leu	Arg	Lys	Leu	Glu	
				155					160					165
Lys	Gln	Ala	Arg	Lys	Ala	Gln	Arg	Glu	Ala	Glu	Glu	Ala	Glu	Arg
				170					175					180
Glu	Glu	Arg	Lys	Arg	Leu	Glu	Ser	Gln	Arg	Glu	Ala	Glu	Trp	Lys
				185					190					195
Lys	Glu	Glu	Glu	Arg	Leu	Arg	Leu	Glu	Glu	Glu	Gln	Lys	Glu	
				200					205					210
Glu	Glu	Arg	Lys	Ala	Arg	Glu	Glu	Gln	Ala	Gln	Arg	Glu	His	Glu
				215					220					225
Glu	Tyr	Leu	Lys	Leu	Lys	Glu	Ala	Phe	Val	Val	Glu	Glu	Gly	
				230					235					240
Val	Gly	Glu	Thr	Met	Thr	Glu	Glu	Gln	Ser	Gln	Ser	Phe	Leu	Thr
				245					250					255
Glu	Phe	Ile	Asn	Tyr	Ile	Lys	Gln	Ser	Lys	Val	Val	Leu	Glu	
				260					265					270
Asp	Leu	Ala	Ser	Gln	Val	Gly	Leu	Arg	Thr	Gln	Asp	Thr	Ile	Asn
				275					280					285
Arg	Ile	Gln	Asp	Leu	Leu	Ala	Glu	Gly	Thr	Ile	Thr	Gly	Val	Ile
				290					295					300
Asp	Asp	Arg	Gly	Lys	Phe	Ile	Tyr	Ile	Thr	Pro	Glu	Glu	Leu	Ala
				305					310					315
Ala	Val	Ala	Asn	Phe	Ile	Arg	Gln	Arg	Gly	Arg	Val	Ser	Ile	Ala
				320					325					330
Glu	Leu	Ala	Gln	Ala	Ser	Asn	Ser	Leu	Ile	Ala	Trp	Gly	Arg	Glu
				335					340					345
Ser	Pro	Ala	Gln	Ala	Pro	Ala								
				350										

<210> 57
<211> 216
<212> PRT
<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 2345712

<400> 57

Tyr	Asp	Pro	Ile	Gly	Phe	Gly	Leu	Ser	Trp	Glu	Ala	Gly	Arg	Ile
1										10				15
Ile	Gly	Trp	Gly	Lys	Pro	Thr	Arg	Gly	Arg	Gly	Arg	Gly	Gly	Ser
				20						25				30
Leu	Ser	Thr	Arg	Gly	Arg	Gly	Ser	Glu	Val	Pro	Asp	Ser	Ala	His
				35						40				45
Leu	Ala	Pro	Thr	Pro	Leu	Phe	Ser	Glu	Ser	Gly	Cys	Cys	Gly	Leu
				50						55				60
Arg	Ser	Arg	Phe	Leu	Thr	Asp	Cys	Lys	Met	Glu	Glu	Gly	Asn	
				65						70				75
Leu	Gly	Gly	Leu	Ile	Lys	Met	Val	His	Leu	Leu	Val	Leu	Ser	Gly
				80						85				90
Ala	Trp	Gly	Met	Gln	Met	Trp	Val	Thr	Phe	Val	Ser	Gly	Phe	Leu
				95						100				105
Leu	Phe	Arg	Ser	Leu	Pro	Arg	His	Thr	Phe	Gly	Leu	Val	Gln	Ser
				110						115				120
Lys	Leu	Phe	Pro	Phe	Tyr	Phe	His	Ile	Ser	Met	Gly	Cys	Ala	Phe
				125						130				135
Ile	Asn	Leu	Cys	Ile	Leu	Ala	Ser	Gln	His	Ala	Trp	Ala	Gln	Leu
				140						145				150
Thr	Phe	Trp	Glu	Ala	Ser	Gln	Leu	Tyr	Leu	Leu	Phe	Leu	Ser	Leu
				155						160				165
Thr	Leu	Ala	Thr	Val	Asn	Ala	Arg	Trp	Leu	Glu	Pro	Arg	Thr	Thr
				170						175				180
Ala	Ala	Met	Trp	Ala	Leu	Gln	Thr	Val	Glu	Lys	Glu	Arg	Gly	Leu
				185						190				195
Gly	Gly	Glu	Val	Pro	Gly	Ser	His	Gln	Gly	Ser	Asp	Pro	Tyr	Arg
				200						205				210
Gln	Leu	Arg	Glu	Lys	Asp									
				215										

<210> 58

<211> 292

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No.: 1810320

<400> 58

Met	Ala	Gln	Pro	Pro	Pro	Asp	Val	Glu	Gly	Asp	Asp	Cys	Leu	Pro
1									10					15
Ala	Tyr	Arg	His	Leu	Phe	Cys	Pro	Asp	Leu	Leu	Arg	Asp	Lys	Val
										25				30
Ala	Phe	Ile	Thr	Gly	Gly	Gly	Ser	Gly	Ile	Gly	Phe	Arg	Ile	Ala
										40				45
Glu	Ile	Phe	Met	Arg	His	Gly	Cys	His	Thr	Val	Ile	Ala	Ser	Arg
										55				60
Ser	Leu	Pro	Arg	Val	Leu	Thr	Ala	Ala	Arg	Lys	Leu	Ala	Gly	Ala
										70				75
Thr	Gly	Arg	Arg	Cys	Leu	Pro	Leu	Ser	Met	Asp	Val	Arg	Ala	Pro
										85				90
Pro	Ala	Val	Met	Ala	Ala	Val	Asp	Gln	Ala	Leu	Lys	Glu	Phe	Gly
										100				105
Arg	Ile	Asp	Ile	Leu	Ile	Asn	Cys	Ala	Ala	Gly	Asn	Phe	Leu	Cys
										115				120
Pro	Ala	Gly	Ala	Leu	Ser	Phe	Asn	Ala	Phe	Lys	Thr	Val	Met	Asp
										130				135
Ile	Asp	Thr	Ser	Gly	Thr	Phe	Asn	Val	Ser	Arg	Val	Leu	Tyr	Glu

Lys	Phe	Phe	Arg	Asp	His	Gly	Gly	Val	Ile	Val	Asn	Ile	Thr	Ala
140									145					150
155									160					165
Thr	Leu	Gly	Asn	Arg	Gly	Gln	Ala	Leu	Gln	Val	His	Ala	Gly	Ser
170									175					180
Ala	Lys	Ala	Ala	Val	Asp	Ala	Met	Thr	Arg	His	Leu	Ala	Val	Glu
185									190					195
Trp	Gly	Pro	Gln	Asn	Ile	Arg	Val.	Asn	Ser	Leu	Ala	Pro	Gly	Pro
200									205					210
Ile	Ser	Gly	Thr	Glu	Gly	Leu	Arg	Arg	Leu	Gly	Gly	Pro	Gln	Ala
215									220					225
Ser	Leu	Ser	Thr	Lys	Val	Thr	Ala	Ser	Pro	Leu	Gln	Arg	Leu	Gly
230									235					240
Asn	Lys	Thr	Glu	Ile	Ala	His	Ser	Val	Leu	Tyr	Leu	Ala	Ser	Pro
245									250					255
Leu	Ala	Ser	Tyr	Val	Thr	Gly	Ala	Val	Leu	Val	Ala	Asp	Gly	Gly
260									265					270
Ala	Trp	Leu	Thr	Phe	Pro	Asn	Gly	Val	Lys	Gly	Leu	Pro	Asp	Phe
275									280					285
Ala	Ser	Phe	Ser	Ala	Lys	Leu								
				290										

<210> 59
<211> 158
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 964996

<400> 59

Glu	Gly	Gly	Pro	Ser	Trp	Thr	Arg	Glu	Arg	Thr	Leu	Val	Ala	Val
1								10						15
Lys	Pro	Asp	Gly	Val	Gln	Arg	Arg	Leu	Val	Gly	Asp	Val	Ile	Gln
								20						30
Arg	Phe	Glu	Arg	Arg	Gly	Phe	Thr	Leu	Val	Gly	Met	Lys	Met	Leu
								35						45
Gln	Ala	Pro	Glu	Ser	Val	Leu	Ala	Glu	His	Tyr	Gln	Asp	Leu	Arg
								50						60
Arg	Lys	Pro	Phe	Tyr	Pro	Ala	Leu	Ile	Arg	Tyr	Met	Ser	Ser	Gly
								65						75
Pro	Val	Val	Ala	Met	Val	Trp	Glu	Gly	Tyr	Asn	Val	Val	Arg	Ala
								80						90
Ser	Arg	Ala	Met	Ile	Gly	His	Thr	Asp	Ser	Ala	Glu	Ala	Ala	Pro
								95						105
Gly	Thr	Ile	Arg	Gly	Tyr	Phe	Ser	Val	His	Ile	Ser	Arg	Asn	Val
								110						120
Ile	His	Ala	Ser	Asp	Ser	Val	Glu	Gly	Ala	Gln	Arg	Glu	Ile	Gln
								125						135
Leu	Trp	Phe	Gln	Ser	Ser	Glu	Leu	Val	Ser	Trp	Ala	Asp	Gly	Gly
								140						150
Gln	His	Ser	Ser	Ile	His	Pro	Ala							
				155										

<210> 60
<211> 559
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701884305H1

<400> 60

gaaaacctaa acgcgcgtgc gcttcttcca cgccacggaa accgtgcagg cctgggtgg 60
 tctccaaagt gactgaacaa tgcagaagga cagtggccca ctggttcctt tacattatta 120
 tggttcggc tatgcggccc tggtggtac tggtgggatt attggctatg caaaagcagg 180
 tagtgtgccg tccctggctg ctggactctt cttggggggc ctggcaggcc tgggtgccta 240
 ccagctgtct caggacccca ggaacgtgtg ggtttccta gctacgtctg ggactttggc 300
 tggcattatg gggatgagat tctacaactc tggaaattt atgcctgcag gttgatcgc 360
 gggagccagt ttgctgtatc ttgccaact tgacttagt atgttgagtt caccatcc 420
 gtagtagcca tagtcctgcg tggcctatc atgagttgac actctccatg cttcacatt 480
 accacgctga agagataaga acagcaaaga cctacactga gcacatggag gcaagacgt 540
 gtttactata gtgaccgtc 559

<210> 61

<211> 326

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701607951H1

<400> 61

tggttgggtg tgttcttact ttgcggattt taccaccctg gaattgttcc gtacgcgcag 60
 ggcgcggggc gctctccgt gcaactctcg ctgagctagc ggactgccc cctctctaaa 120
 acgtcctgtt actgcgggtt cgggagtgga aacctaaacg cgcgtgcgt tctccacgc 180
 cacggaaacc gtgcaggcct ggtgtggctt ccaaagtgac tgaacaatgc agaaggacag 240
 tggcccaactg gttcctttac attattatgg ttccgttat gcggccctgg tggctactgg 300
 tgggattatt ggctatgcaa aagcag 326

<210> 62

<211> 333

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701644253H1

<400> 62

aacgtcctgt aactgcgggtt ccgggagtggtt aaacctaaac ggcgcgtgc tttttccac 60
 gccacggaaa accgtgcagg cctngtgtgg tcctcanagt gactgaacaa tgcagaagga 120
 cagtggccca ctggntcctt tacattatta tggttcggc tatgcggccc tggtgctac 180
 tggtggttatttggctatg caaaagcagg tagtgcggc tccctggctg ctggactctt 240
 ctttgggggc ctggcaggcc tgggtgccta ccagctgtct caggacccca ggaacgtgtg 300
 gttttccta gctacgnctg ggactttggc tgg 333

<210> 63

<211> 318

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701513151H1

<400> 63

cttactttgc ggattttacc accctggaat tggtccgtac ggcgcangngc gcggggctct 60
 cccgtgcact ctctgtgtgg ctgcggact gcccgcctt ctaaaacgtc ctgttaactgc 120
 ggttccggga gtggaaacctt aaacgcgcgt ggcgttctt caccgcacgg aaaccgtgca 180
 ggcctgggtgt ggtctccaaa gtgactgaac aatgcagaag gacagtggcc cactggttcc 240
 ttacattat tatgggttcg gctatgcggc cttgggtggct actgggtggaa ttatggcta 300

tgcaaaaagca ggttagtgt

318

<210> 64
<211> 315
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701652337H1

<400> 64

cagcncaggc ctccgggctc cagctccggc gttgggttca ggcctgggt ggtctccaaa 60
gtgactgaac aatgcagaag gacagtggcc cactggttcc ttacattat tatggtttcg 120
gctatgcggc cctgggtggc actgggtggaa ttattggcta tgcaaaaagca ggttagtgtgc 180
cgccccggc tgctggactc ttctttgggg gcctggcagg cctgggtgcc taccagctgt 240
ctcaggaccc caggaacgtg tgggactttcc tagctacgtc tgggactttg gctggcatat 300
ggggatgaga ttcta 315

<210> 65
<211> 313
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701562183H1

<400> 65

ggtctccaaa gtgactgaac aatgcagaag gacagtggcc cactggttcc ttacattat 60
tatggtttcg gctatgcggc cctgggtggc actgggtggaa ttattggcta tgcaaaaagca 120
ggttagtgtgc cgccccggc tgctggactc ttctttgggg gcctggcagg cctgggtgcc 180
taccagctgt ctcaggaccc caggaacgtg tgggactttcc tagctacgtc tgggactttg 240
gctggcatta tggggatgag attctacaac tctggaaat ttatgcotgc aggtttgatc 300
gcgggancat ttt 313

<210> 66
<211> 304
<212> DNA
<213> Rattus norvegicus

<220> misc_feature
<223> Incyte ID No.: 700227356H1

<400> 66

cgccgtcgtc ctccagcgcga ggcctccggg ctccagctcc ggtgtgggt gcaggcctgg 60
tgtggctcc aaagtgtactg aacaatgcag aaggacagtg gcccactggg tccttacat 120
tattatggt tcggctatgc gcccctggg gctactggg ggattattgg ctatgcaaaa 180
gcaggttagtgc tgccgtccct ggctgtggc ctcttctttg gggcctggc aggctgggt 240
gcctaccagc tgtctcaggc ccccaggaac gtgtgggtt tcctagctac gtctggact 300
ttgg 304

<210> 67
<211> 327
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701649802H1

<400> 67

ctccgggtgtt ggggcagggc ctgggtgtt ctc当地aaatgt actgaacaat gcagaaggac 60
agtggaccac tggttccta cattattatg gtgcggcta tgc当地ccctg gtggctactg 120
gtgggattat tgncttca aaagcaggta gtgtgc当地tgc cctggctgtt ggactcttct 180
ttggggccct ggcaggcctg ggtgc当地tacc agctgtctca ggaccccagg aacgtgtggg 240
tttccttagc tacgtctggg actttggctg gcattatggg gatgagattc tacaactctg 300
gaaaattat gcctgcagtt tgatcgc 327

<210> 68

<211> 305

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700226414H1

<400> 68

gccgtcgccc tccagcncaag gcctccgggc tccagctccg gtgttgggtg caggcctgg 60
gtgggtctcca aagtgtactg acaatgcaga aggacagtgg cccactgtt cctttacatt 120
attatgtttt cggctatgcg gccc当地gggtgg ctactgggtg gattatggc tatgcaaag 180
caggtagtgt ggc当地ccctg getgtggac tttctttgg gggc当地ggca ggctgggtg 240
cctaccatgt gtctcaggac cccaggaaat gtgggttttctgctacgt ctgggacttg 300
gctgg 305

<210> 69

<211> 295

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700275094H1

<400> 69

tcctccagcn caggcncccg ggctccagct cc当地gttgg gtgc当地ccct gggtgtgtct 60
ccaaatgtac tgaacaatgc agaaggacag tggcccactg gttc当地ttac attattatgg 120
tttc当地gttat gccc当地ctgg tggctactgg tggattatt ggctatgcaa aacaggtag 180
tgtgc当地gtcc ctggctgctg gactcttctt tggggccct ggc当地ccctg ggtgc当地tacc 240
agctgtctca ggaccccagg aacgtgtggg tttc当地tagc tacgtctggg atttg 295

<210> 70

<211> 301

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700226425H1

<400> 70

cctgacctct gttc当地gtgc tccc当地ccgtc gtc当地ccagc gcaggc当地cc gggtccaggc 60
tccgggtttt ggtgc当地ggc tgggtggc tccaaatgt ctgaacaatg cagaaggaca 120
gtggccact gttc当地tta cattattatg gttc当地ggc tgc当地ccctg gtggctactg 180
gtgggattat tggctatgca aaagcaggta gtgtgc当地tgc cctggctgtt ggactcttct 240
ttggggccct ggc当地ccctg ggtgc当地tacc agctgtctca ggaccccagg aacgtgtggg 300
t 301

<210> 71

<211> 282

<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700275207H1

<400> 71

tcctccagcg caggcctccg ggctccagct ccgggtgttgg gtgcaggcct ggtgtggct 60
ccaaagtgc tgaacaatgc agaaggacag tgcccactg gttccttac attattatgg 120
tttcggctat gcggccctgg tggctactgg tggattatt ggctatgcaa aagcaggtag 180
tgtccgtcc ctggctgctg gactcttctt tgccccctg gcaggcctgg gtgcctacca 240
gctgtctcag gaccaggaa acgtgtgggt ttccctagct ac 282

<210> 72
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701507568H1

<400> 72

cggcgtcgta ctccagcgca ggcctccggg ctccagctcc ggtgtgggt gcaggcctgg 60
tgtggcttcc aaagtgc tggctatgc aacaatgc aaggacagtg gcccactgg tcccttacat 120
tattatggtt tcggctatgc ggccctggg gctactggg ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgtggaa ctcttctttt ggggcctggc aggctgggt 240
gcctaccaggc tgtctcagga ccccaggaa acgtgtgggtt tc 282

<210> 73
<211> 281
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700300118H1

<400> 73

cggcgtcgta ctccagcgca ggcctccggg ctccagctcc ggtgtgggt gcaggcctgg 60
tgtggcttcc aaagtgc tggctatgc aacaatgc aaggacagtg gcccactgg tcccttacat 120
tattatggtt tcggctatgc ggccctggg gctactggg ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgtggaa ctcttctttt ggggcctggc aggctgggt 240
gcctaccaggc tgtctcagga ccccaggaa acgtgtgggtt t 281

<210> 74
<211> 292
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700301710H1

<400> 74

cctgnacctc tgccctgtg ctccggcgt cgccctccag cgccaggcctc cgggctccag 60
ctccgggtt ggggcaggc ctgggtgtt ctccaaatgc actgaacaat gcagaaggac 120
agtggccac tggcttccctt acattattat ggttccgtt atgcggccct ggtggctact 180
ggtgggatata tggctatgc aaaaggcaggta agtgcgtccgt ccctggctgc tggactctc 240
tttgggggcc tggcaggcctc ggggcctac cagctgtctc aggaccccg ga 292

<210> 75
<211> 289
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700064344H1

<400> 75

cagcgcaggc ctccgggctc cagctccgt gttgggtgtg ttcttacttt gcggattta 60
ccaccctgga attgttccgt acgcgcaggc gcgcgggcgc tctccgtgc actctctgct 120
gagctagcgg actgcccgc tctctaaaac gtccctgtaac tgcggttccg ggagtggaaa 180
cctaaacgcg cgtgcgccttc ttccacgcca cgaaaaccgt gcaggcctgg tgtggtctcc 240
aaagtatgatga acatgcagaa ggacantggc ccactggttc ttanatatt 289

<210> 76
<211> 276
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701423273H1

<400> 76

agcgcaggcc tcagggctcc agctccgtg ttgggtgcag gcctgggtgn gtctccaaag 60
tgactgaaca atgcagaagg acagtggccc actggttctt tacattatt atggttcgg 120
ctatgcggcc ctgggtggcta ctgggtggat tattggctat gcaaaagcag gtatgtgcc 180
gtccctggct gctggactct tctttgggg cctggcagge ctgggtgcct accagctgtc 240
tcaggacccc aggaacgtgt gggtttcct agctac 276

<210> 77
<211> 293
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700225847H1

<400> 77

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<210> 78
<211> 274
<212> DNA
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<220>
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<223> Incyte ID No.: 701462776H1

<400> 78

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ttacattatt atggttcgg ctatcgccc ctggtggtta ctgggtggat tattggctat 180
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<210> 79
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<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700916803H1

<400> 79

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tctccctgtc actctctgtc gagctagcg actgcccggc tctctaaaac gtctgtAAC 180
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<210> 80
<211> 280
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 80

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ttatggtttc ggctatgcgg ccctgggtgc tactgggtggg attattggct atgaaaaac 180
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<210> 81
<211> 299
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701646690H1

<400> 81

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ngccctgggt gctactgggt ggattattgg ctatgcaaaa ncaggttagtgc tgccgtccct 180
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<210> 82
<211> 286
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701624261H1

<400> 82

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ggtgggattt ttggctatgc aaaagcaggt agtgtgccgt ccctggctgc nngacttcc 180
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<210> 83

<211> 266

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700912920H1

<400> 83

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tggactcttc tttgggggccc tggcaggccct gggtgcctac cagctgtctc aggaccccag 180
gaacgtgtgg gttttccctag ctacgtctgg gactttggct ggcatttatgg ggatgagatt 240
ctacaactct gggaaattta tgccctg 266

<210> 84

<211> 262

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701482566H1

<400> 84

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atgagattct acaactctgg gaaattttagt cctgcagggtt tgatcgccgg agccagttt 180
ctgatggttt ccaaacttgg acttagttagt ttgagttcac cccatccgta gtagccatag 240
ccctgcgtgg gctcatgtat ag 262

<210> 85

<211> 285

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700270272H1

<400> 85

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ctggttccctt tacattatta tggttcggc tatgcggccc tggtggtac tggtggtt 180
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<210> 86

<211> 268

<212> DNA

<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700628520H1

<400> 86

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tcggctatgc ggccctgggt gctactgggt ggattattgg ctatgcaaaaa gcaggttagtg 180
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tgtctcagga cccccaggaac gtgtgggt 268

<210> 87
<211> 269
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700534975H1

<400> 87

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ttacattant atggtttccg ctatgcggcc ctgggtggcta ctgggtggat tattggctat 180
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ctgggtgcct accagctgtc tcaggaccc 269

<210> 88
<211> 262
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700176004H1

<400> 88

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caggacccca ggaacgtgtg gggtttccctt gctacgtctg ggactttggc tggcattatg 180
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ttgctgtatgg ttgccaact tg 262

<210> 89
<211> 349
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701609236H1

<400> 89

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ttcttccacg ccacggaaac cgtgcaggcc tgggtgtggtc tccaaagtga ctgaaacaatg 180
cagaaggaca gtggcccact gggttcccttta cattattatg gtttcggcta tgccggccctg 240
gtggctactg gtggatattt ggctatgcaa aagcagtatg tgccgtccct ggctgctgaa 300
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<210> 90
<211> 263
<212> DNA
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<220>
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<223> Incyte ID No.: 701473437H1

<400> 90

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tcaggacccc aggaacgtgt ggg 263

<210> 91
<211> 303
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701606089H1

<400> 91

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gga 303

<210> 92
<211> 273
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701736525H2

<400> 92

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<210> 93
<211> 262
<212> DNA
<213> Rattus norvegicus

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<223> Incyte ID No.: 701532848H1

<400> 93

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<210> 94
<211> 247
<212> DNA
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<400> 94

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cagtggccca ctggttcctt tacattatta tggttcggc tatgcggccc tggggctac 180
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<210> 95
<211> 284
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 95

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ctggtggtcta ctgggtggat tattggctat gcaaaaaggcag gtagtgtgcc gtcctggct 240
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<210> 96
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 96

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<210> 97
<211> 281
<212> DNA
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<223> Incyte ID No.: 701481465H1

<400> 97

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<210> 98

<211> 265

<212> DNA

<213> Rattus norvegicus

<220>

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<223> Incyte ID No.: 701308467H1

<400> 98

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<210> 99

<211> 291

<212> DNA

<213> Rattus norvegicus

<220>

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<223> Incyte ID No.: 701564368H1

<400> 99

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<211> 271

<212> DNA

<213> Rattus norvegicus

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<210> 101

<211> 255

<212> DNA

<213> Rattus norvegicus

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<400> 101

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<210> 102
<211> 297
<212> DNA
<213> Rattus norvegicus

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atggttgcca aacttggact tagtatgtt agttcacccc atccgtagta gccatag 297

<210> 103
<211> 261
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 103

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<210> 104
<211> 312
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 104

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<210> 105
<211> 241
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 105

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g 241

<210> 106
<211> 268
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
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<400> 106

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<210> 107
<211> 318
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 107

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<210> 108
<211> 255
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701293154H1

<400> 108

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tttcctagct acgtctggga ctttggcttg cattatgggg atgagattct acaactctgg 180
gaaatttatg cctgcagggt tgatcgccgg agccagttt ctgatggttt ccaaacttgg 240
attatgtatgt tgagg 255

<210> 109
<211> 254
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701298824H1

<400> 109

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tgccgcctg gctgctggac tcttctttgg gggcctgcag nctgggtgcc taccagctgt 240
ctcaggaccc agga 254

<210> 110
<211> 294
<212> DNA
<213> Rattus norvegicus

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<223> Incyte ID No.: 700524204H1

<400> 110

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<210> 111
<211> 289
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700067537H1

<400> 111

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<210> 112
<211> 276
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 112

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cctgcagnct ggtgcctacc agctgctcg cgtngg 276

<210> 113

<211> 254

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700532493H1

<400> 113

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tttgcgtatg gttgccaaac ttggacttag tatgttgagt tcaaaaaatc cgttagtagcc 180
atagcctgc gtgggctcat gatgagttgc atctccagtc ctctacatta ccacgctgaa 240
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<210> 114

<211> 282

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700523302H1

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aaagtactg aacaatgcag aaggacagtg gcccactgg tcccttanat aatnatggtc 120
gggtanangn ncccgnnnnng nnaagggggn atttgnnnnt acgnaagagc ngntagtgtg 180
ccgtccctgg ctgctggact cttcttggg ggcctggcag gcctgggtgc ctaccagctg 240
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<210> 115

<211> 256

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701242719H1

<400> 115

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gcagaaggac agtggccac tggcctttt acattattat ggtttcggt atgcggccct 180
ggnggctact ggtgggattta ttggctatca aaagcaggta gtgtgccccc ctggctgtgg 240
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<210> 116

<211> 244

<212> DNA

<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701226025H1

<400> 116

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tgcctaccag ctgctcagga ccccagaac gtgtgggtt tccttagctac gtctggact 180
ttgctggcat tatggggatg agattctaca actctggaa atttatcctg caggtttgat 240
cgcg 244

<210> 117
<211> 262
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701293276H1

<400> 117

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gaacaatgca gaaggacagt ggcactgg ttcccttaca ttattatggt ttccgctatc 180
ggcccttgggt ggctactggg gggattattg gctatgaaa agcaggtagt gtgccgtccc 240
tggctgtgga ctctctntgn gg 262

<210> 118
<211> 261
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700493358H1

<400> 118

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tatgaaaaag caggtatgtt gccgtccctg gctgctggac tttctttgg ggncntggca 180
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<210> 119
<211> 265
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700533285H1

<400> 119

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tggcccaactg gttcctttac attattatgg ttccggctat gggccctgg tggctactgg 180
tgggattattt ggtatgcaaa agcaggtagt gtgccgtccc tggctgtgg actttcttt 240
ggggccctgg caggcctggg tgcct 265

<210> 120
<211> 247
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700920823H1

<400> 120

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gtgactgaac aatgcagaag gacagtggcc cactggttcc ttacattat tatggttcg 180
gctatgcggc cctgggtggct actgggtggaa ttatgttat gcaaaaagcag gtagtctgcc 240
gctccct 247

<210> 121
<211> 263
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700627607H1

<400> 121

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tgaacaatgc agaaggacaa tggcccaactg gtccctttac attattatgg ttccggctat 180
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ggcnanctcg nnccgaggng nnc 263

<210> 122
<211> 265
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700437944H2

<400> 122

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tgnntgggatt attggctatn caaaagcagg tagtgtncgg tccctggctg ctggactctt 180
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ngttttctta agctacntct gggac 265

<210> 123
<211> 343
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701582848H1

<400> 123

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gcgggagcca nttgctata gttgccaact tngacttagt atgttgagtn caccccatcc 180

gtagtagcat ancctgcgtg ggctcagatg agtnacactc tccaggcctc cacatttacc 240
aggctgaaga gtaagacagc aaagactaca tgagcacntg aggnaaacgt ggtntatat 300
gacgttcaag acgcgatgt gactcagact ncntgctcat cg 343

<210> 124
<211> 241
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701305531H1

<400> 124

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caggcctccg ggctccagct ccggagttgg gtgcaggcct ggnngtgnct ccaaagtgac 120
tgaacaatgc agaaggacag tggcccactg gttcctttac attattatgg attcggctat 180
gcggccctgg tggctactgg tggattattg gctatcaaaa gcaggagtgt ccggccctgct 240
g 241

<210> 125
<211> 155
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700916103H1

<400> 125

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tttacattat tatggttcg gctatgcggc cctgg 155

<210> 126
<211> 185
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701294764H1

<400> 126

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attattatgg ttteggctat gggccctgg aggcnactgg gggnatattg gctatncaa 180
agcgg 185

<210> 127
<211> 125
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700066710H1

<400> 127

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gtgtgggtt tcctagctac gtctggact ttggctggca ttatgggat gagattctac 120
aactc 125

<210> 128
<211> 266
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701471559H1

<400> 128

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gacactctcc agtcctctac attaccacgc tgaagagata agaacagcaa agacctacac 180
tgagcacatg gaggcagaaga cgtggttact atagtgaccg ttcagagntg gcgagtgtct 240
gacctcagag ctcacactgc cttcat 266

<210> 129
<211> 208
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700325006H1

<400> 129

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ttatgcctgc aggttnatc gcggnccancc agttgnntg atggttgcca aacttggact 180
tagtangntn anttcacccca ntgcgcgtc 208

<210> 130
<211> 263
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701258479H1

<400> 130

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gacctctctt ccacaggtgc aggccctggc tgnctccaa agtgaactgaa caatgcagaa 180
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actggngna ttattggcta tgc 263

<210> 131
<211> 258
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700627187H1

<400> 131

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taggatgttgc agttcacccc atcccgaggat agccatagtc ctgcgtggc tcatgtatgag 120
ttgacactct ccagtccctcc acattaccac gctgaagaga taagaacacgc aaagacctac 180
actgagcaca tggaggcgaa gacgtggta ctatagtgac cgttcagaga cggcgagtgt 240
ctgactcaga gctcacac 258

<210> 132

<211> 272

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 701246066H1

<400> 132

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Ottaccacgct gaagagatan gaacagcaaaa gacctacact gagcacatgg aggcgaagac 180
gtggttacta tagtgaccgt tcagagacgg cgagtgtctg acctcagacg tcacactgct 240
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<210> 133

<211> 253

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700594190H1

<400> 133

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acgtggtac tatagtgacc gttcagagac ggcgagtgtc tgacctcaga gctcacactg 180
ccttcatgcg gcttggctt gtgtcatgat gtctcgactc tctgtactac tacataaagg 240
gttaaatgt tgg 253

<210> 134

<211> 267

<212> DNA

<213> Rattus norvegicus

<220>

<221> misc_feature

<223> Incyte ID No.: 700627108H1

<400> 134

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tgnntgaac actctccagt cagtccagat naacgnccgt gntagagatn aagaccagcn 180
aagacctaca ctgagcacca tggaggcgaa gacgtggta ctataagtga ccgttcagag 240
acggcgngtg tntggatcan agatcca 267

<210> 135

<211> 650

<212> DNA

<213> Rattus norvegicus

<220>
 <223> RnAUG.conN

<400> 135

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tctccgtgc actctctgct gagctagcg actgcccgc tctctaaaac gtctgttaac 180
tgcggttccg ggagtggaaa cctaaacgcg cgtgcgcctc ttccacgcca cgaaaaccgt 240
gcaggcctgg tgtggtctcc aaagtgactg aacaatgcag aaggacagt gcccactgg 300
tccttacat tattatggt tcggctatgc gggcctggtg gctactggg ggattattgg 360
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gtctgggact ttggctggca ttatgggat gagattctac aactctggga aatttatgcc 540
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gagttcaccc catccgtagt agccatagcc ctgcgtggc tcatgatgag 650
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<210> 136
 <211> 114
 <212> PRT
 <213> Rattus norvegicus

<220>
 <223> RnAUG.conP

<400> 136

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									25					30
Ala	Lys	Ala	Gly	Ser	Val	Pro	Ser	Leu	Ala	Ala	Gly	Leu	Phe	Phe
									35					45
Gly	Gly	Leu	Ala	Gly	Leu	Gly	Ala	Tyr	Gln	Leu	Ser	Gln	Asp	Pro
									50					60
Arg	Asn	Val	Trp	Val	Phe	Leu	Ala	Thr	Ser	Gly	Thr	Leu	Ala	Gly
									65					75
Ile	Met	Gly	Met	Arg	Phe	Tyr	Asn	Ser	Gly	Lys	Phe	Met	Pro	Ala
									80					90
Gly	Leu	Ile	Ala	Gly	Ala	Ser	Leu	Leu	Met	Val	Ala	Lys	Leu	Gly
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Leu	Ser	Met	Leu	Ser	Ser	Pro	His	Pro						
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<210> 137
 <211> 223
 <212> DNA
 <213> Homo sapiens

<220>
 <221> misc_feature
 <223> Incyte ID No.: 746355H1

<400> 137

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<210> 138
 <211> 243
 <212> DNA
 <213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1294663H1

<400> 138

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ggagttcgta ttttgatgac atctgatttt cagaagtcat gttccagctt ggactcatga 120
aggattaaaa atctgcatct tccactatTT tcaatgtatt aagagaaaata agtgcagcat 180
ttttgcacatct gacatTTTAC ctaaaaaaaaaa aaagacacca aatttggcgg aggggtggaa 240
aat 0 243